1 Topology-driven analysis of protein-protein interaction

2 networks detects functional genetic modules regulating

3 reproductive capacity

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Abstract

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- 11 Understanding the genetic regulation of organ structure is a fundamental problem in
- developmental biology. Here, we use egg-producing structures of insect ovaries, called
- ovarioles, to deduce systems-level gene regulatory relationships from quantitative functional
- 14 genetic analysis. We previously showed that Hippo signalling, a conserved regulator of animal
- organ size, regulates ovariole number in *Drosophila melanogaster*. To comprehensively
- 16 determine how Hippo signalling interacts with other pathways in this regulation, we screened all
- 17 known signalling pathway genes, and identified Hpo-dependent and Hpo-independent signalling
- 18 requirements. Network analysis of known protein-protein interactions among screen results
- 19 identified independent gene regulatory modules regulating one or both of ovariole number and
- 20 egg laying. These modules predict involvement of previously uncharacterised genes with higher
- 21 accuracy than the original candidate screen. This shows that network analysis combining
- 22 functional genetic and large-scale interaction data can predict function of novel genes regulating
- 23 development.

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- 25 **Keywords**: *Drosophila melanogaster*, Reproduction, Ovariole, Ovary, Egg laying, Topology,
- 26 Network analysis, Interactome, Hippo signalling.

Introduction

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The final shape and size of an organ is critical to organismal function and viability. Defects in human organ morphology cause a multitude of pathologies, including cancers, organ hypertrophies and atrophies (e.g. Yang and Xu. 2011). It is thus critical to understand the regulatory mechanisms underlying the stereotypic shape and size of organs. To this end, assessing the genetic regulation of size is significantly facilitated by using quantifiable changes in organ size and shape. The Drosophila melanogaster female reproductive system is a useful paradigm to study quantitative anatomical traits. In these organs, the effects of multiple genes and the environment combine to produce a quantitative phenotype: a species-specific average number of eggproducing ovarian tubes called ovarioles. Fruit fly ovaries can contain as few as one and as many as 50 ovarioles per ovary, depending on the species (Kambysellis and Heed, 1971; King, 1970; Markow et al., 2009; Sarikaya et al., 2019), with each ovariole capable of producing eggs. Ovariole number, therefore, may affect the reproductive fitness of *Drosophila* species by determining the potential of an adult female to produce eggs (Klepsatel et al., 2013b; R'kha et al., 1997). While ovariole number within a species can vary across temperatures (Azevedo et al., 1996), altitudinal and latitudinal clines (Capy et al., 1994; David and Bocquet, 1975), under constant environmental conditions ovariole number is highly stereotypic (Capy et al., 1993; Klepsatel et al., 2013a; R'Kha et al., 1991; R'kha et al., 1997). The reproducibility of ovariole number thus indicates a strong genetic component (Sarikaya et al., 2019). Genome wide association studies and quantitative trait locus mapping have demonstrated that the ovariole number is a highly polygenic trait (Bergland et al., 2008; Lobell et al., 2017; Orgogozo et al., 2006; Wayne et al., 2001; Wayne et al., 1997; Wayne and McIntyre, 2002). In contrast, functional genetic studies have identified only a small number of genes whose activity regulates ovariole number (discussed below). Thus, the complexity of the genetic regulation of this important trait remains largely unknown. The determination of ovariole number in *D. melanogaster* occurs during late larval and pupal development (King et al., 1968). Each ovariole in the adult fly arises from a single primordial structure called a terminal filament (TF), which forms in the late third instar larval ovary (Godt and Laski, 1995) by convergent extension (Keller, 2006) of the terminal filament cells (TFCs) (Godt and Laski, 1995; Sahut-Barnola et al., 1996). TFCs are first specified from an anterior

population of somatic cells in the larval ovary by the expression of transcription factors including Bric-à-brac 1/2 (bric-à-brac 1/2; bab1/2) and Engrailed (engrailed; en) (Godt and Laski, 1995) (Sahut-Barnola et al., 1995). Initially a loosely arranged group in the anterior of the larval ovary. TFCs undergo morphogenetic movements to give rise to the ordered columns of cells that are TFs. Cell intercalation during convergent extension is dependent on the actin regulators Cofilin (twinstar) and the large Maf factor Traffic Jam (traffic jam; tj), and on E-cadherin dependent adhesion (Chen et al., 2001; Godt and Laski, 1995). Regulation of ovariole number is thus largely dependent on the specification of the TFCs and their rearrangement into TFs (Sarikaya and Extavour, 2015). We previously showed that the regulation of both TFC and TF number is dependent on the Hippo signalling pathway (Sarikaya and Extavour, 2015), a pan-metazoan regulator of organ and tissue size (Hilman and Gat, 2011; Sebe-Pedros et al., 2012). At the core of the Hippo kinase cascade are two protein kinases, Hippo (hippo; hpo) and Warts (warts), which prevent the nuclear localisation of the transcriptional co-activator Yorkie (yorkie; yki). Yki and the transcription factor Scalloped (scalloped) together initiate the transcription of multiple gene targets, including those that promote cell proliferation and survival. In the D. melanogaster larval ovary, loss of Hpo in the somatic cells causes an increase in nuclear Yki, leading to an increase in TFCs, TFs, ovariole number and egg laying in adults (Sarikaya and Extavour, 2015). Production of fertile eggs from a stereotypic number of ovarioles requires a spatially and temporally coordinated interplay of signalling between the somatic and germ line cells of the ovary. Thus, signalling amongst somatic and germ line cells in the larval ovary is crucial to all stages of ovarian development (Ables and Drummond-Barbosa, 2017; Gilboa, 2015; Green II et al., 2011; LaFever and Drummond-Barbosa, 2005; LaFever et al., 2010; Sarikaya and Extavour, 2015). For instance, disruptions in insulin or Tor signalling affect both somatic and germ line cell proliferation (Gancz and Gilboa, 2013; Green II and Extavour, 2012; Hsu and Drummond-Barbosa, 2009; LaFever and Drummond-Barbosa, 2005; LaFever et al., 2010; Sarikava et al., 2012). Similarly, ecdysone pulses from the prothoracic gland regulate the timely differentiation

of the primordial germ cells (PGCs) and the somatic TFCs (Gancz et al., 2011; Hodin and

line to somatic cells by differentially regulating proliferation of both cell types (Gancz et al.,

Riddiford, 1998, 2000b). Both Hpo and ecdysone signalling also control the proportion of germ

2011; Sarikaya and Extavour, 2015).

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Although it is clear that genes function together in regulatory networks (Gonzalez and Kann, 2012), determining how the few genes functionally verified as required for ovariole development and function, work together to coordinate ovariole number and ovarian function more generally. is a challenge because most genes or pathways have been considered individually. An alternative approach that is less often applied to animal developmental genetics, is a systems biology representation of complex biological systems as networks (Barabási and Albert, 1999; Watts and Strogatz, 1998). Protein-protein interaction (PPI) networks are such an example (Albert and Barabási, 2002). The availability of high throughput molecular biology datasets from, for example, yeast two-hybrid, protein CHiP and microarrays has allowed for the emergence of large scale interaction networks representing both functional and physical molecular interactions (Barabási and Oltvai, 2004; Berger et al., 2007; Giot et al., 2003; Gonzalez and Kann, 2012). With ample evidence that signalling in the ovary can affect ovarian development, but few genes functionally verified to date, we aimed to identify novel regulators of ovariole development by functionally testing all known members of all characterized D. melanogaster signalling pathways. We used tissue-specific RNAi to systematically knock down 463 genes in the larval ovary, and looked for modifiers of the hpo loss of function egg laying and ovariole number phenotypes. To analyse the results of this phenotypic analysis, we used topology-driven network analysis to identify genetic modules regulating these phenotypes, thus generating hypotheses about the relationships between these modules. With this systems biology approach, we identify not only signalling pathway genes, but also previously untested genes that affect these reproductive traits. Functional testing showed that these novel genes affect ovariole number and/or egg laying, providing us with a novel in silico method to identify target genes that affect ovarian development and function. We use these findings to propose putative developmental regulatory modules underlying one or both of ovariole formation and egg laying.

Results

- An RNAi modifier screen for signalling pathway involvement in ovariole
- 124 number

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- To systematically ascertain the function of signalling pathway genes and their interactions with
- Hippo signalling in the development of the *D. melanogaster* ovary, we first curated a list of all

known and predicted signalling genes (Gramates et al., 2016; Kanehisa et al., 2010; Mbodj et al., 2013). We identified 475 genes belonging to the 14 developmental signalling pathways characterised in D. melanogaster (Table 1; Table S1), and obtained UAS:RNAi lines for 463 of these genes from the Vienna Drosophila RNAi centre (VDRC) or the TRiP collections at the Bloomington Drosophila Stock centre (BDSC) (all D. melanogaster genetic lines used are listed in Methods). We previously showed that reducing the levels of hpo in the somatic cells of the larval ovary using traffic jam Gal4 (tj:Gal4) driving hpo[RNAi] increased both ovariole number and egg laying of adult female flies (Sarikaya and Extavour, 2015). To identify genes that modify these phenotypes, we used ti: Gal4 to drive simultaneous hpo[RNAi] and RNAi against a signalling candidate gene, and quantified the phenotypic change (Figure 1a-d). We observed that on driving two copies of hpo[RNAi] using tj:Gal4, we obtained a further increase in both egg laying and ovariole number (Figure 1e). This indicates that ovaries have further potential to increase ovariole number and egg laying beyond the increase induced by ti:Gal4 driving one copy of hpo[RNAi], and that tj:Gal4 can drive the expression of two RNAi constructs, indicating that our screen could identify both enhancers and suppressors of the ti:Gal4>hpo[RNAi] phenotype. We proceeded to identify modifiers of the ti:Gal4>hpo[RNAi] phenotype by crossing males of each of the 463 candidate genes RNAis individually with tj:Gal4>hpo[RNAi] females, and performing three phenotypic screens on the offspring. In the first screen (Figure 1a), we measured egg laying of three F1 female offspring (tj:Gal4>hpo[RNAi], signalling candidate[RNAi]) over 5 days. To address batch variation (Figure S1), we standardized egg laying measurements by calculating the Z scores (Z_{gene} = number of standard deviations from the mean) for each candidate line relative to its batch controls. 190 genes had an egg laying $|Z_{gene}|$ below 1. Previous studies have shown that the egg laying of newly eclosed adult mated females correlates with ovariole number during the first five days (Klepsatel et al., 2013b). We therefore eliminated these 190 genes from subsequent screening, because the change in egg laying was so modest that we considered these candidates were unlikely to show changes in ovariole number when compared to controls. In the second screen (Figure 1b), we measured egg laying in a wild-type background (tj>signalling candidate[RNAi]) for the 273 remaining candidate genes. For the third screen (Figure 1c), we quantified the ovariole number of ti:Gal4>hpo[RNAi], signalling candidate[RNAi]

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F1 adult females for the same 273 candidate genes. To choose candidates from the second and third screens for further study, we wished to account for the fact that the two screens had different effective numbers of data points. This was because egg laying data were obtained from individual vials of three females over five days, while ovariole numbers were obtained from 20 ovaries from ten females (see methods). We therefore selected the 67 genes with a $|Z_{gene}|$ above two for ovariole number (Figure 1c, 1d; Table 2), and the 49 genes with a more conservative $|Z_{gene}|$ above five for egg laying (Figure 1a, 1b, 1d; Table 2), for a total of 116 positive candidates for subsequent analyses.

Ovariole number is weakly correlated with egg laying

Standardization of the results from the three screens using Z scores allowed us to compare the effects of individual genes on one or both of egg laying and ovariole number. We performed a pairwise comparison of the Z_{gene} values for all combinations of screens, and considered genes with $|Z_{gene}|$ values that were above the thresholds set for the phenotype in each screen (above two for ovariole number, above five for egg laying; green dots in Figure 2a-c. Across all three screens, loss of function of our positive candidates yielded reductions in ovariole number and egg laying more commonly than increases (Figure 2a-c). Comparing the $|Z_{gene}|$ values of egg laying and ovariole number of tj:Gal4>hpo[RNAi], signalling candidate[RNAi] adult females revealed that genes that caused a change in egg laying did not always similarly affect ovariole number, and vice versa (Figure 2a). We therefore hypothesise that egg laying and ovariole number may be regulated by genetically separable mechanisms. This hypothesis notwithstanding, we observed a weak but statistically significant correlation between egg laying and ovariole number (p=1e10⁻⁵; Figure 2d), and this correlation was most significant in adult females that had a drastic reduction in both phenotypes (Figure 2a).

No single signaling pathway dominates regulation of ovariole number or

egg laying

We found that at least some genes from all tested signalling pathways could affect both egg laying and ovariole number (Figure 3). To determine if some pathway(s) appeared to play a more important role than others in these processes, we asked whether any of our screens were

enriched for genes from a specific signalling pathway. To measure enrichment, we compared the distribution of individual pathway genes among the positive candidates in each screen to a bootstrapped null distribution of pathway genes among a group of the same number of genes randomly selected from our curated list of 463 signalling genes (Figure 3a). Involvement of a pathway in the regulation of a phenotype would be reflected in a difference between the representation of pathway genes in an experimentally derived list and a randomly selected group of signalling genes. We found that rather than only one or a few pathways showing functional evidence of regulating ovariole number or egg laying, nearly all pathways affected both phenotypes (Figure 3a). We further tested this result by calculating the hypergeometric pvalue for the enrichment of each signaling pathway, in each of the three groups of genes. Consistent with the results of the bootstrapping approach (Figure 3a), we found that most pathway members were not significantly enriched for egg laving or ovariole number phenotypes (Figure 3b). The absence of significant enrichment of any specific pathway is not simply attributable to the pool of genes that were screened, because our experimental manipulations of ovariole number and egg laying did cause a change in the distribution of signalling pathway members (Figure S2a). Instead, both phenotypes appeared to be regulated by members of most or all signalling pathways (Figure 3; Figure S2). The only two exceptions to this trend were a greater than twofold enrichment of (1) genes from the Notch signalling pathway in the regulation of ovariole number (p-value < 0.05, pink bar in Figure 3a, b), and (2) members of the Hedgehog (Hh) signaling pathway in the regulation of Hippo-dependent egg laying (p-value < 0.05, brown bar in Figure 3a, 3b; Figure S3). In summation, our analyses of the enrichment of signalling pathways within the different screens indicated that both ovariole number and egg laying are regulated by genes from nearly all described animal signalling pathways (Figure 3a), rather than being dominated by any single pathway. Comparing the results of the Egg Laying screens performed in a wild type background (Figure

Comparing the results of the Egg Laying screens performed in a wild type background (Figure 1b) or in a hpo[RNAi] background (Figure 1a), revealed that most of the genes that met a threshold of $|Z_{gene}| > 5$ in one screen, did not meet that threshold in the other screen (Figure 2c). This result suggests the existence of both Hippo-dependent and Hippo-independent mechanisms of regulation of egg laying. The interpretations of separable Hippo-dependent and independent regulation of egg laying, and of the separable regulation of ovariole number and egg laying, was supported by the results of the network analysis described in the following section.

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Centrality of genes in the ovarian protein-protein interaction networks can predict the likelihood of loss of function phenotypic effects

The finding that these reproductive traits were regulated by the genes of all signalling pathways led us to consider the broader topology of putative gene regulatory networks in the analysis of our data. Previously characterized genes in the ovary are often pleiotropic and can regulate both ovariole number and egg laying (Gilboa, 2015; Sarikaya and Extavour, 2015). As with proteins in a linear pathway, proteins in a protein-protein interaction (PPI) network are more likely to function in conjunction with genes that are connected to them within the network (e.g. Ideker and Sharan, 2008; Jeong et al., 2001). Centrality is one measure of the connectedness of a gene in the PPI and can be used to identify the most important functional centres within a protein network (Hahn and Kern, 2005; Ma'ayan, 2011). Most centrality measures use path length, which is a measure of the number of other proteins required to link any two proteins in the network. Here we used four commonly used metrics to quantify gene centrality, each measuring slightly different properties (Jalili et al., 2016; Koschutzki and Schreiber, 2008). (1) Degree centrality is proportional to the number of proteins that a given protein directly interacts with. (2) Betweenness centrality measures the number of shortest paths amongst all the shortest paths between all pairs of proteins that require passing through a particular protein. (3) Closeness centrality measures the average shortest path that connects a given protein to all other proteins in the network. (4) Eigenvector centrality is a measure of the closeness of a given protein to other highly connected proteins within the network.

We hypothesised that if the candidate genes we identified in our screen as playing roles in ovarian function worked together as a PPI network, then the degree of centrality of a gene might be an indicator of function. To test this hypothesis, we calculated the four centrality measures, described above, for all genes within the *D. melanogaster* PPI (Figure S4). We then rank ordered only the genes tested in each screen by their score for each centrality measure, and asked whether their rank order correlated with the results of the screen, plotting these results as a receiver operating characteristic (ROC) curve. Positive correlations between centrality (a continuous variable) and phenotype (a binary variable: above or below the $|Z_{gene}|$ threshold) are reflected in an area under the curve (AUC) of more than 0.5. We found that the higher the centrality score, the greater the likelihood that a gene had $|Z_{gene}|$ values above our threshold for effects on ovariole number and egg laying (Figure 4a; Table S3). This supports the premise that the positive candidates identified in our screen function together as a network in the regulation

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of either ovariole number or egg laying. Interestingly, while the centrality of genes did predict whether a gene would affect our phenotypes of interest, it could only weakly predict the strength of that effect (Figure S5).

Genes regulating egg laying and ovariole number regulation form nonrandom interaction modules

The centrality analyses above suggested that the genes implicated in ovariole number and egglaying displayed characteristics of a functional network. PPI networks can often be further sorted into a collection of sub-networks. A sub-network is a smaller selection of proteins from the PPI. Examples of such sub-networks could be proteins within the same subcellular organelle (Foster et al., 2006) or genes that are expressed at the same time (Spellman et al., 1998), thus making them likely to function together (Srinivasan et al., 2007). A module is a sub-network that can perform regulatory functions independent of other sub-networks, and has key measurable features (Barabási and Oltvai, 2004; Hartwell et al., 1999; Ravasz et al., 2002; Yook et al., 2004). We therefore asked if our sub-networks consisting of genes that showed similar mutant phenotypes might constitute such functional modules. To determine whether genes that were implicated in regulation of ovariole number and egg laying interacted with each other in specific groups more than would be expected by chance, we created four lists of genes, called "seed" lists, based on their individual phenotypic effects based on our screen results: (1) the core seed list, including genes positive in all three screens (Figure 4b); (2) the egg laying seed list, including genes positive in the wild type background egg-laving screen (Figure 1b: Figure 4c): (3) the hpo[RNAi] egg laying seed list, including genes positive in the hpo[RNAi] background egg laying screen (Figure 1a; Figure 4c); and (4) the hpo[RNAi] ovariole seed list, including genes positive in the hpo[RNAi] background ovariole number screen (Figure 1c; Figure 4c). Interestingly, the core seed list, comprising genes that affected all three measured phenotypes, only consisted of genes that caused a reduction in both ovariole number and egg laying (Figure 4b).

Based on published molecular interactions, putative functional modules of genes can be predicted by algorithms that use either the shortest path method (Bromberg et al., 2008) or the Steiner Tree approach (Huang and Fraenkel, 2009). Such methods identify and predict

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functional connections between the seed proteins, as well as additional nodes (proteins or genes) that have not been experimentally tested within the given parameters, but are known to interact with the seed genes in the PPI (Albert and Albert, 2004)((Yu et al., 2006). This process results in a predicted module, and subsequent functional genetic testing of this module can confirm its functionality. Given its recent success in predicting gene modules, we used the previously published Seed Connector Algorithm (SCA), a member of the Steiner Tree algorithm family (Wang et al., 2017; Wang and Loscalzo, 2018), to identify putative functional modules formed by genes that had similar phenotypic effects in our screens (Figure 4b, 4c). The SCA connects seed genes and previously untested novel genes (connectors) to each other using a known PPI network, producing the largest possible connected putative module given the data. We compiled a PPI network consisting of all described interactions between *D. melanogaster* proteins, from the combination of publicly available PPI studies in the DroID database (see Methods). Using this PPI network and the aforementioned four seed lists, we applied a custom python implementation of the SCA (Methods: 04 Seed-Connector.ipynb) to build and extract the largest possible (given our PPI) connected putative modules that regulate egg laying and ovariole number. This SCA method yielded four putative modules, one for each seed list, which we refer to as the core module (Figure 5b), hpo[RNAi] Egg Laying module (Figure S6a), Egg Laying module (Figure S6b), and hpo[RNAi] Ovariole Number Module (Figure S6c) respectively. Each of the modules contained seed genes, which had been functionally evaluated in our screens (green and red circles in Figure 5), as well as connector genes, which were genes newly predicted as regulators of these phenotypes (green and red triangles in Figure 5). We then asked whether these four putative modules were more connected than we would expect by chance; in other words, we formally tested them for modularity. Meeting our criteria for modularity would suggest that the genes in these modules operated together as functional sub-networks within the *Drosophila* PPI. We defined our modularity test using four commonly measured network metrics: (1) Largest Connected Component (LCC) (the number of proteins connected together by at least one interaction), (2) network density (the relative number of edges as compared to the theoretical maximum), (3) total number of edges, and (4) average shortest path (average of the minimum distances connecting any two proteins). We considered a group of genes to form a module if they showed higher LCC, higher network density, more edges, and shorter average shortest path length than a random, bootstrapped selection of the same number of genes from the PPI.

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To determine whether these criteria would correctly identify genes of the same signaling pathway, which are known to function together, as a module, we measured these four parameters in the original set of genes used in this study (Table 1). We found that all the genes of a given signalling pathway were identified as a module based on these parameters (Figure S7a). We then used this approach to test the modularity of the four phenotypic sub-networks, as compared to 1000 bootstrapped "control sub-networks" consisting of a group of the same number of genes as contained in the sub-network, but chosen randomly from among the candidate genes from our initial screen list (Table 1). We found that the four predicted phenotypic modules showed a significantly increased Largest Connected Component (LCC) value, network density, number of edges and decreased average shortest path (Figure S7b), compared to our "control module". This result indicates that these sub-networks are likely to function as modules within the PPI, to regulate one or both of ovariole number or egg laying.

Low edge densities between modules indicates genetically separable mechanisms of ovariole number and egg laying

Our network analysis identified four highly connected networks of genes that regulate two distinct developmental processes, together with or independently of Hippo signalling activity: ovariole number determination, which occurs primarily during larval development, and egg laying, which takes place in adult life (Figure 5). We wished to assess the degree to which there were any shared genetic components between the four modules. To understand potential interactions between the modules in the regulation of both ovariole number and egg laying, we constructed a composite network of all genes in each of the four modules (Figure 5b; Figure S6), which we refer to as the "meta network" (Figure 7a). We then grouped the genes based on their phenotypic effects as measured in the three screens, resulting in seven sub-networks (I-VII in Figure 7a). We then asked if these sub-networks were as connected to each other, as were the genes within each of the sub-networks. To do this, we used an edge density map, which reflects the number of interactions between the genes within a sub-network and between each of the sub-networks (Figure 7b).

This analysis yielded three principal findings. First, edge densities between the three subnetworks corresponding to the three scored phenotypes (I, II and III in Figure 7a) were very low (Figure 7b). This indicates that genes in each of these sub-networks function as largely independent networks, rather than interacting substantially with any genes in the other non-core sub-networks. Second, the core sub-network (IV in Figure 7a) displayed a higher edge density with the other three sub-networks (I, II and III in Figure 7a) than any of them did with each other (Figure 7b). Consistent with the definition of core module genes as regulating all three scored reproductive phenotypes, this result suggests that the core module genes, in contrast to those from the other three sub-networks, may share substantial functional interactions with genes of the other sub-networks. Finally, three small additional sub-networks emerged from this analysis (V, VI and VII in Figure 7a), suggesting small functional networks of genes that work together to regulate two of the three scored phenotypes. In sum, this meta network analysis supports the hypothesis of three largely independent genetic networks that regulate Hippo-dependent ovariole number, Hippo-dependent egg laying, and Hippo-independent egg laying. Moreover, each of these genetically separable networks included genes in multiple signalling pathways (Figure 7c).

Network analysis predicts novel genes involved in egg laying and ovariole number

The four predicted phenotypic modules produced by the SCA approach included connector genes that were not included in our original screen, and thus had not been tested for possible effects on our phenotypes of interest (triangles in Figure 5b; Figure S6). Given that prior work in human disease models showed that predicted disease modules can correctly predict gene involvement in the relevant diseases (Chen et al., 2006; Gonzalez et al., 2007; Wang et al., 2017; Wang and Loscalzo, 2018), we asked whether our deployment of the SCA had likewise successfully predicted novel, functionally important genes in each module. To do this, we measured the effects of knocking down each the connector genes (triangles in Figure 5b and Figure S6) on ovariole number and egg laying, using *UAS:RNAi* lines for each connector, driven by *tj:Gal4*.

Of the ten predicted novel connectors in the core module, loss of function of several of these had significant effects on at least one of the three scored phenotypes. Five affected ovariole

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number two affected Hpo-dependent egg laying, and one affected Hpo-independent egg laying. 382 However, only one of them significantly altered all three scored phenotypes (Figure 6a; Table 383 S4). The predicted connector genes from two of the other three phenotypic modules showed high 385 positive prediction rates. RNAi against seven out of 18 of the hpo[RNAi] Egg Laving module 386 connectors, three out of 11 of the hpo[RNAi] Ovariole Number module connectors, and none of the 11 Egg Laying module connectors, significantly affected the module phenotype (Table S4). Thus, although the Egg Laying module connectors failed to impact this phenotype in our assay, 41.2% and 27.3% of the connectors from the other two modules were correctly predicted (Figure 6b; Table S4). These positive hit rates exceed those obtained in our initial candidate screens, where 59/463 (12.7%) and 67/273 (24.5%) tested genes affected hpo-dependent egg laying and hpo-dependent ovariole number respectively (Figure 6d; Table 2). In sum, taken across all modules (Figure 6c; Table S4, Table S5), this network analysis correctly identified genes regulating all scored reproductive phenotypes, at rates higher than those obtained in the 395 original screen of 463 members of all known signalling pathways. By this measure, testing network-predicted regulatory modules derived from experimentally obtained data was even 397 more successful than testing signalling pathways as a means of identifying novel genes that regulate ovariole number and egg laying.

Discussion

- Identification of regulatory modules for ovariole development and egg
- 402 laying

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- 403 The *D. melanogaster* ovary is a commonly studied model for organogenesis (Chen et al., 2001:
- 404 Godt and Laski, 1995; Lobell et al., 2017; Sarikaya and Extavour, 2015), stem cell maintenance
- 405 (Gilboa, 2015) and interactions of development and ecology (Cohet and David, 1978; Hodin and
- 406 Riddiford, 2000a; Klepsatel et al., 2013a; Sarikaya et al., 2019). Nevertheless, our
- 407 understanding of the genetic mechanisms that regulate these processes remains fragmentary.
- 408 In this paper, we have identified four distinct protein interaction modules that regulate ovariole
- 409 number and egg laying in the *D. melanogaster* ovary. These modules consist of both novel and

previously characterized genes that regulate either ovariole number or egg laying or both, thus enhancing our understanding of the genetic underpinnings of this reproductive system.

Of the four modules, the core module affects both ovariole number and egg laying. The core module contains numerous housekeeping genes, including regulators of transcription, translation and cell division such as *polo* (Llamazares et al., 1991), *cyclin K* (Edwards et al., 1998), *nucleosome assembly protein 1* (Ito et al., 1996) and *eukaryotic translation release factor 1* (Chao et al., 2003). While *polo* and *eukaryotic translation release factor 1* are members of signalling pathways, *cyclin K* and *nucleosome assembly protein 1* are genes predicted by the SCA. Given that the core module largely consists of genes whose loss of function decreases both of these parameters, we hypothesise that these are essential genes for the basic structure and function of the ovaries. Essential genes are more interconnected in a PPI with higher centrality measures (Jeong et al., 2001) and interestingly, we find that the genes in the core module also have higher connectivity than those in the other three modules (Figure S4).

In addition to genes that regulate basic cellular processes, the core module is enriched for the core components of the Hh signalling cascade, namely *patched (ptc)*, *smoothened (smo)* and *costa (cos)* (Lee et al., 2016). However, we find that the loss of Hh ligand, which is expressed in the TF cells in the developing larval ovary (Lai et al., 2017), does not significantly affect either ovariole number or egg laying. Though surprising, ligand-independent activation of Hedgehog signalling has been observed before. For example, in the *Drosophila* eye, loss of either *ptc* or *cos* in clones leads to non-cell autonomous proliferation in wild type cells, as well as growth disadvantages in the mutant tissue (Christiansen et al., 2012). In another example, sufficient intracellular *smo* levels can also activate downstream transcription of Hh pathway targets, showing that Hh itself is not always required to activate the cascade (Jiang et al., 2018).

Development of the larval ovary

The *hpo[RNAi]* Ovariole Number module is composed of genes that affect the Hippo signalling activity-dependent determination of ovariole number during development. Establishment of ovariole number occurs largely during the third instar stage of larval development in *D. melanogaster* (Godt and Laski, 1995; Hodin and Riddiford, 1998; King et al., 1968; Sahut-Barnola et al., 1996). During this period, the TFCs are specified in the anterior of the ovary and undergo rearrangement into stacks of cells called TFs, each of which gives rise to an ovariole (Godt and Laski, 1995; Sahut-Barnola et al., 1995). TF specification requires the expression of

443 engrailed (En) (Bolívar et al., 2006) and the transcription factors Bab1 and Bab2, encoded by 444 the bric-à-brac locus (Couderc et al., 2002; Godt et al., 1993). A third transcription factor, 445 Lmx1a, was recently found to be necessary for the specification of the TFCs (Allbee et al., 446 2018). Our hpo[RNAi] Ovariole Number module identifies numerous additional novel 447 transcription factors including bunched (bun) and retinoblastoma-family protein (rbf), which we 448 hypothesize could also be involved in the specification of ovariole number. bun and rbf have 449 been implicated in the migration (Dobens et al., 2005) and endoreplication (Cayirlioglu et al., 450 2003) of the follicle cells during oogenesis, but have not, to our knowledge, been previously 451 identified as playing a role in the context of larval ovary development. 452 453 The TFCs specified in the larval ovary undergo a process of convergent extension to form TFs. 454 This process of convergent extension requires cell intercalation, and the actin depolymerizing 455 factor Cofilin, encoded by the gene twinstar, is essential to this process (Chen et al., 2001). 456 During intercalation, the cells also dynamically modify their actin cytoskeleton and their 457 expression of E-cadherin (Godt and Laski, 1995). Our hpo[RNAi] Ovariole Number module 458 further identifies Rho1 (Barrett et al., 1997) and Rho kinase (Rok) (Mizuno et al., 1999) as 459 necessary for correct ovariole number. During the extension of the *D. melanogaster* embryonic 460 germ band, a commonly studied model of convergent extension, the localised activation of the 461 actin-myosin network facilitated by Rho1 and Rok is necessary for cell intercalation (Kasza et 462 al., 2014). Given the known roles of Rho1 and Rok as regulators of the actin cytoskeleton 463 (Ridley, 2006), we propose that TF assembly in the ovary requires both these proteins for 464 correct cell intercalation. A third actin cytoskeleton regulator, misshapen (msn), was also 465 identified by our hpo[RNAi] Ovariole Number module. msn encodes a MAP kinase previously 466 shown to regulate the polarisation of the actin cytoskeleton during oogenesis (Lewellyn et al., 467 2013), but has not, to our knowledge, been studied to date in the context of larval ovarian 468 development. 469 470 We propose that the polarity of the somatic cells in the ovary is also necessary for correct larval 471 ovary development, given the presence of the lateral membrane proteins discs large 1 (dlg1) 472 and prickle (pk) in the ovariole module. During the maturation of the TFs during larval 473 development, the TFCs undergo significant cell shape changes, coincident with localised 474 expression of beta-Catenin and actin to the lateral edges of the TFCs (Godt and Laski, 1995). 475 Restriction of the E-cadherin domain in epithelia requires establishment of the basolateral

domain (Harris and Peifer, 2004) and we propose that testing a similar requirement for *dlg1* and *pk* in the larval ovary would be a fruitful avenue for future studies.

Network analysis as a tool in developmental biology

Using a systems biology approach to analyse RNAi screening data has proven fruitful, providing us with new insights into the development and function of the *D. melanogaster* ovary by identifying novel and previously understudied genes that regulate this process. Systematic analysis of the function of single genes in development has been a historical convention and has provided valuable and precise genetic interaction information (Jansen et al., 2002; von Mering et al., 2002). With the advent of genome-wide analysis, however, we can use data from a larger number of genes to predict the identity of additional functionally significant genes with relative ease (Yu et al., 2006). We note that the novel gene prediction rate ranged from as high as 41.2% from the hpo[RNAi] Ovariole Number module to as low as 0% from the Egg Laying module (Figure 6b; Tables S4, S5). We suggest that this may be due to multiple factors. Firstly, the possible incompleteness of the PPI is expected to lead to some areas of the network being sparse or non-existent (von Mering et al., 2002). If the module of interest happens to fall in such regions of the network, prediction algorithms will fail. Secondly, the initial restriction of tested genes to signaling pathway members might have provided a seed list too sparse to usefully predict functional connectors. Finally, it could be the case that "Egg Laving" is such a complex phenotype that its gene regulation cannot be adequately captured within a highly connected network of the type suited for identification by the analyses we have used here. Ovariole number in *D. melanogaster* is the outcome of a discrete developmental process with a clear beginning and end, comprising a specific series of cellular behaviors that take place in the confines of one organ (Godt and Laski, 1995; Hodin and Riddiford, 2000a; Sahut-Barnola et al., 1996). Once established during larval life, ovariole number in Drosophila remains unaltered through to and during adulthood, even if oogenesis within those ovarioles suffers congenital or age-related defects (King, 1970). Because previous work suggested that ovariole number in Drosophila could have at least some predictive relation to egg laying (Cohet and David, 1978; Klepsatel et al., 2013b; Sarikaya and Extavour, 2015), we reasoned that scoring the latter phenotype in a primary screen (Figure 1a) could be an effective way to uncover ovariole number regulators (Figure 1c). While our results showed that this was true in many cases, it was also clear that these two traits can vary independently (Figure 2), highlighting the fact that ovariole number is not the only determinant of egg laying. Egg-laying dynamics, even during the limited

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five day assay used in our study, are likely influenced not just by a single anatomical parameter such as ovariole number, but rather by many biological, biomechanical, hormonal and behavioural processes. Consequently, the functional module we were able to extract from the results of this screen (Figure 1b) might be too coarse to extract novel genes that participate in potentially complex gene interactions regulating egg laying. The predictive rates of the approach we have used here, although encouraging, are likely limited by the degree of noise in the high throughput data used to generate the PPI (Li et al., 2010), the sparseness of the PPI, and the degree of misidentification of protein interactions (Zhang et al., 2017). Addressing one or more of these parameters could improve the outcomes of future network predictions from developmental genetics data. For example, the problem of sparseness, which is a paucity of high confidence detectable interactions relative to all biologically relevant interactions, has been addressed in other studies by using an "Interolog PPI" (Matthews et al., 2001) in place of an organism-specific PPI. The Interolog PPI combines known interactions from multiple organisms, and has been used successfully to identify, for example, gene modules relevant in squamous carcinoma, based on a starting dataset of microarray data on differentially expressed genes between cancer cells and the surrounding tissue (Wachi et al., 2005). Future studies applying the Interolog PPI to the outcomes of genetic screens for developmental processes of interest could potentially overcome the problem of sparseness, as well as the biases towards proteins that are more heavily studied and thus better represented in organism-specific PPIs.

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Tables

529 Table 1

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Signalling pathway	Number of genes in screen
EGF	45
FGF	25
FOXO	67
Нірро	60
JAK/STAT	31
JNK	28
MAPK	29
Notch	48
SHH	54
TGF B	52
Toll	36
VEGF	17
Wnt	125
mTOR	36

Table 1: Number of candidate genes tested in each signalling pathway. Candidate genes are grouped by their reported roles in one or more signalling pathways based on published literature. Genes in this list are not necessarily unique to a single pathway, but may function in more than one signalling pathway. The list of specific genes per pathway that were included in the screen for functional analysis (Figure 1) is found in Table S1.

Table 2

Egg Laying Screens	hpo[RNAi] Egg Laying (Figure 1a)	Egg Laying (Figure 1b)	Ovariole Number Screen	hpo[RNAi] Ovariole Number (Figure 1c)
RNAi stocks unavailable	12	0	RNAi stocks unavailable	0
Primary filter ($ Z_{gene} < 1$)	190	N/A	Primary filter ($ Z_{gene} < 1$)	N/A
No effect (-5 < $ Z_{gene} $ < 5)	214	224	No effect (-2< $ Z_{gene} $ < 2)	206
Negative effect ($Z_{gene} < -5$)	48	44	Negative effect ($Z_{gene} < -2$)	54
Positive effect ($Z_{gene} > 5$)	11	5	Positive effect ($Z_{gene} > 2$)	13
Total	475	273	Total	273

Table 2: Results of the three functional genetic screens. Number of genes tested in each screen and cumulative results. "Negative effect" corresponds to a reduction in eggs laid or number of ovarioles below the Z score (Z_{gene}) threshold for each phenotype. "Positive effect" indicates an increase above the set Z_{gene} thresholds. Z_{gene} thresholds for each category in each screen are indicated in brackets. The primary filter of $|Z_{gene}| < 1$ was applied only to the hpo[RNAi] Egg Laying screen shown in Figure 1a. The list of specific genes that exceeded our chosen Z_{gene} thresholds for each scored phenotype (Figure 1), and were therefore considered to have a positive or negative effect on the phenotype, is found in Table S1. The 12 genes for which RNAis stocks were unavailable at the time of testing are listed in Table S2.

548 Figure 1

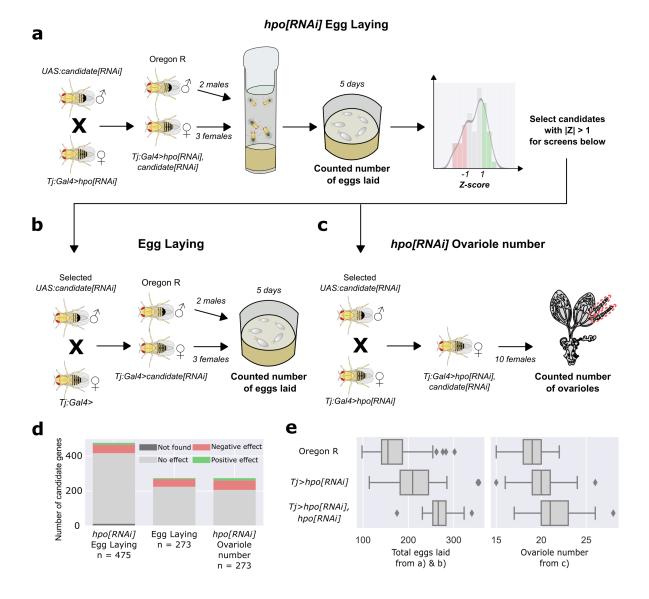


Figure 1. Screen methodology. a,b,c) Diagrammatic representation of screen workflow. **d**) Distributions of results of genes in the three screens. n = number of genes tested in each screen (see also Table 2). **e**) Total eggs laid by three female flies over five days (left panel) and ovariole number (right panel) of Oregon R (top row), *tj:Gal4* driving one copy of *UAS:hpo[RNAi]* (middle row), and *tj:Gal4* driving two copies of *UAS:hpo[RNAi]* (bottom row), showing that, the previously reported *tj:Gal4>hpo[RNAi]* ovariole number and egg laying phenotypes (Sarikaya and Extavour, 2015) can be modified by further UAS:RNAi-mediated gene knockdown.

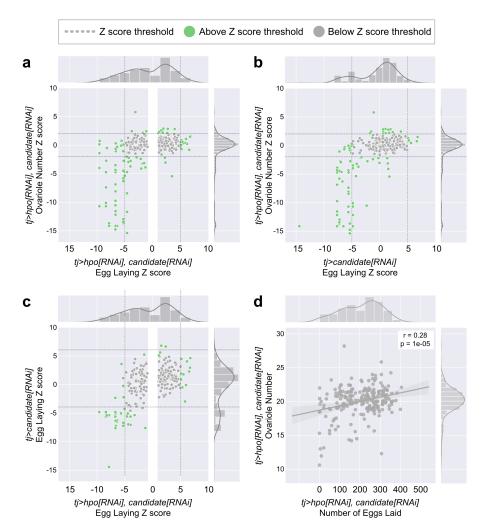


Figure 2. Relationship between Egg Laying and Ovariole Number phenotypes generated in the screens. a) Scatter plots of the Z score for each gene (Z_{gene}) of egg laying versus the ovariole number of adult tj>hpo[RNAi], candidate[RNAi] females. b) Scatter plots of the Z score for each gene (Z_{gene}) of egg laying of adult tj>candidate[RNAi] females versus the ovariole number of adult tj>hpo[RNAi], candidate[RNAi] females. c) Scatter plots of the Z score for each gene (Z_{gene}) of egg laying of adult tj>candidate[RNAi] females versus egg laying of adult tj>hpo[RNAi], candidate[RNAi] females. In a, b and c, bar graphs on the top and right sides of each panel show the distribution of genes in each axis of the adjacent scatter plots . Green dots = genes that meet the Z_{gene} threshold for the indicated phenotype. Grey dots = genes that do not meet the Z_{gene} threshold for the indicated phenotype. Dark grey dotted lines = thresholds for each phenotype: $|Z_{gene}|$ > 5 for Egg Laying and $|Z_{gene}|$ > 2 for Ovariole Number. In a and c, the white vertical bar removes all genes in the tj>hpo[RNAi], candidate[RNAi] with a $|Z_{gene}|$ <1 for egg laying. These genes were not measured in the other two conditions and are therefore not represented in the scatter plots. d) Correlation between non-zero Ovariole Number and Egg Laying values.

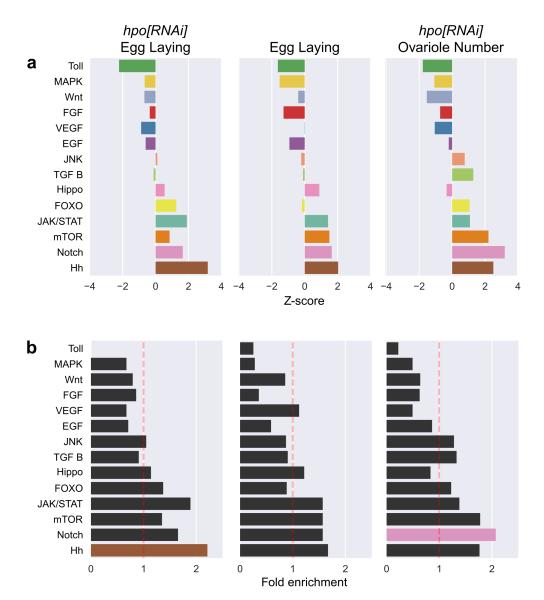


Figure 3. Enrichment of genes of individual signalling pathways among the experimentally obtained positive candidates of each screen. a) Depletion/enrichment analysis to identify over- or under- represented members of individual signalling pathways among positive candidates of each screen. Positive Z scores represent an enrichment, and negative Z scores represent depletion, of genes of a pathway among those genes that experimentally affected the phenotype Enrichment and depletion are defined relative to a null distribution of the expected number of members of a signalling pathway among a group containing the same number of randomly selected signalling genes. b) Fold enrichment and hypergeometric p-value calculation to identify over- or under-representation of the genes of a pathway in each screen. Significantly enriched pathways (colored bars: brown = Hedgehog; pink = Notch) are defined by having a hypergeometric p-value less than 0.05.

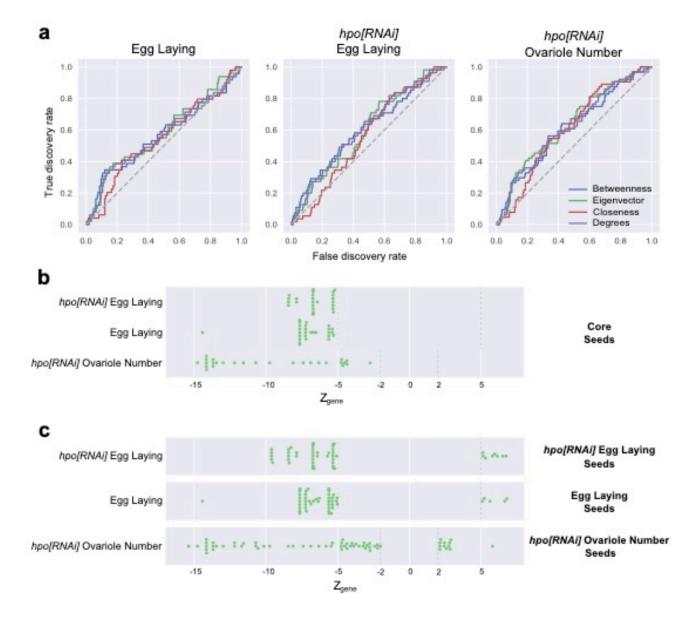


Figure 4. Screened genes function as a network. a) Receiver operating characteristic (ROC) curves of genes ordered by rank for each of four network centrality metrics (Betweenness centrality, Eigenvector centrality, Closeness centrality and Degree centrality) versus a binary outcome (above or below Z score threshold) for each of the three screens. For each screen and metric, the Area Under the Curve (AUC) is > 0.5 (Table S3). **b)** Genes whose $|Z_{gene}|$ value was above the threshold (green dots; Table 2) in all three screens were assigned to the Core seed list. **c)** Genes whose $|Z_{gene}|$ value was above the threshold (green dots; Table 2) in each screen were assigned to the corresponding seed list.

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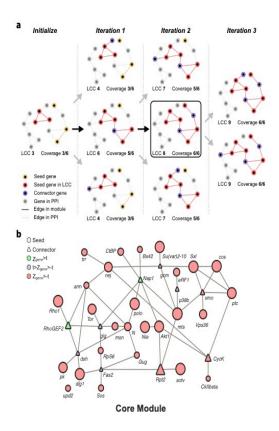


Figure 5. Representation of Seed Connector Algorithm and output. a) Schematic representation of the seed connector algorithm. The algorithm initializes by creating a subnetwork of seed genes from the PPI, and computes the Largest Connected Component (LCC) and coverage (number of genes from the seed set in the LCC). At each iteration, genes in the direct neighborhood of the LCC (distance = 1) are added one at a time to the seed set, and the coverage and LCC are recomputed. This process is repeated for each gene in the direct neighborhood, each time restarting from the seed set of the preceding iteration. If any gene outside the seed set but in the direct neighbourhood is found to maximize coverage while minimizing the LCC, it is added to the seed set as a connector gene. Black arrows indicate the path taken by the algorithm for which the criteria of maximal coverage and minimal LCC are met; such a path would be used to proceed to the subsequent iteration. Grey arrows indicate paths that fail to meet these criteria; such paths would be disregarded. The iteration repeats until the coverage cannot be increased; in this schematic example, this state is achieved in iteration 3. b) The Core Module generated by the Seed Connector Algorithm (SCA) based on the results of the genetic screens (Figure 1a-c). The size of the shapes indicate the relative Z_{gene} score of the gene. Circles indicate seed genes (functionally tested in the screen; Table 2; Table S1) while triangles are connector genes (novel predicted genes; Tables S1, S4, S5). Green = genes with a positive Z_{gene} score above the threshold; red = genes with a negative Z_{gene} score above the threshold; grey = genes with Z_{gene} values below the threshold.

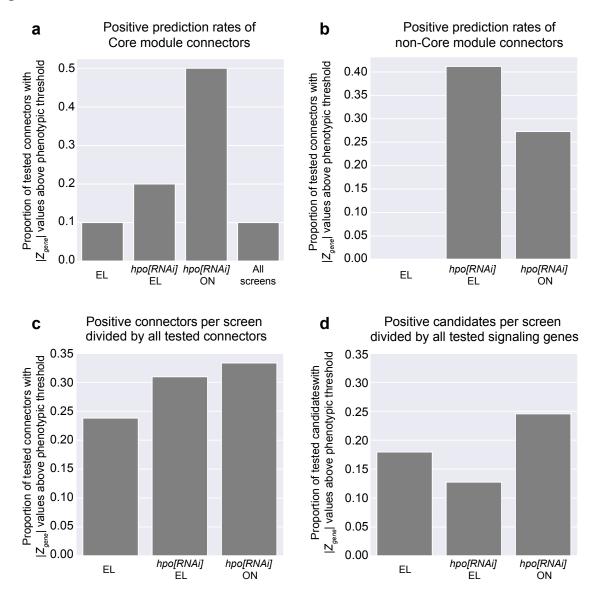


Figure 6. Positive prediction rates of the connector genes in each of the four modules. a) Proportion of core module connector genes with $|Z_{gene}|$ above the threshold in each of the three screens. The "All phenotypes" category includes the genes with $|Z_{gene}|$ above the threshold in all three screens. b) Proportion of tested connector genes in each of the three modules with $|Z_{gene}|$ above the threshold within their corresponding screen. c) Proportion of all unique connector genes predicted by all four modules with $|Z_{gene}|$ above the respective threshold in any of the three screens. d) Proportion of positive candidate genes emerging from the three original signalling candidate screens with $|Z_{gene}|$ above the threshold relative to the total number of genes tested in each screen.

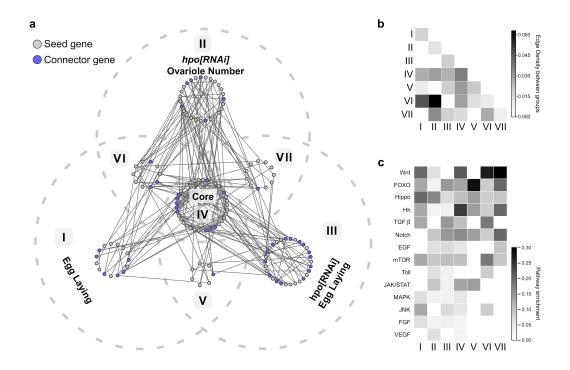
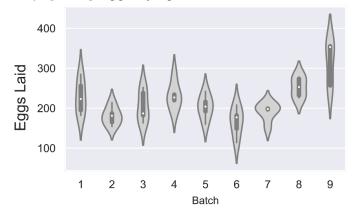


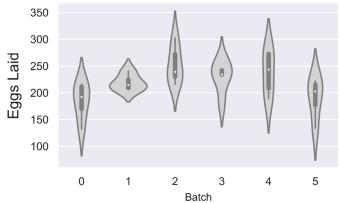
Figure 7. Phenotypically separable sub-networks formed by analysis of the combined genes from all modules. The meta network is generated by the union of the genes in the four phenotypic modules: <code>hpo[RNAi]</code> Egg Laying (Figure S6a), Egg Laying (Figure S6b), <code>hpo[RNAi]</code> Ovariole Number (Figure S6c) and Core (Figure 5b). a) The meta network is represented as a Venn diagram, in which each grey dotted outline represents the screen in which a given gene was identified as affecting the scored phenotype. Within each sub-network, grey circles indicate seed genes, and blue circles indicate connector genes. A single gene, <code>sloppy paired 1</code>, was a seed in the Egg Laying module and also a connector in the <code>hpo[RNAi]</code> Egg Laying module; it fell within sub-network VII in the meta network, and is marked as a seed (grey) in this figure. Solid grey lines indicate interactions between genes in the meta network from the PPI. b) Edge densities between the seven sub-networks of the meta-network. c) Relative enrichment of screened members of the 14 tested developmental signalling pathways within the seven sub-networks of the meta-network.

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a. hpo[RNAi] Egg Laying



b. Egg Laying



c. hpo[RNAi] Ovariole Number

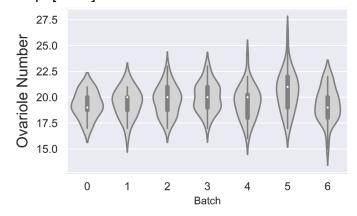


Figure S1. Violin plots of egg laying and ovariole number of controls in each screen batch. a) Distribution of number of eggs laid by five replicates of three *tj:Gal4>hpo[RNAi]* females over five days for each batch. **b)** Distribution of number of eggs laid by five replicates of three *tj:Gal4* females over five days for each batch. **c)** Distribution of number of ovarioles per ovary in 20 ovaries from ten *tj:Gal4>hpo[RNAi]* females in each batch.

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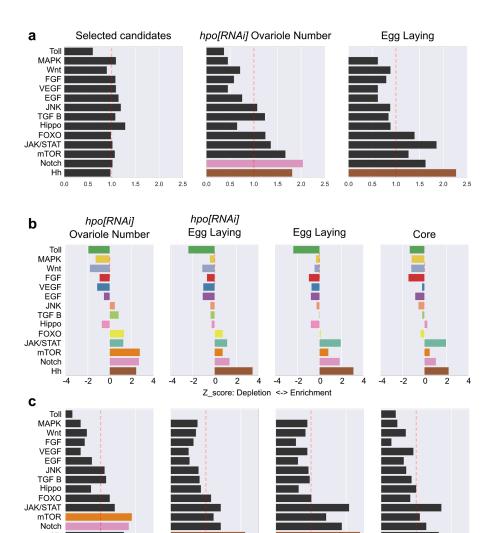


Figure S2. Biological aspects of the network modules. a) Enrichment/depletion analysis of the 273 signalling pathway genes above the threshold $|Z_{gene}| > 1$ (Figure 1a) against all signalling candidates. We also measured the enrichment/depletion of positive signalling candidate genes in the hpo[RNAi] Ovariole (Figure 1c) and Egg Laying (Figure 1b) screens from the 273 genes tested in those screens. **b)** Signalling pathway depletion enrichment analysis. For each module, a null distribution of the expected number of members of a signalling pathway from a group of the same number of randomly selected signalling pathway genes was calculated. The Z score from the expected distribution was then calculated. Negative Z scores represent a depletion, while positive Z scores represent an enrichment. No single pathway is enriched in any of those modules. **c)** Fold enrichment and hypergeometric p-value calculation for each pathway in the four modules. Pathway members in color (orange = mTor; brown = Hedgehog; pink = Notch) have a p-value < 0.05.

Fold enrichment

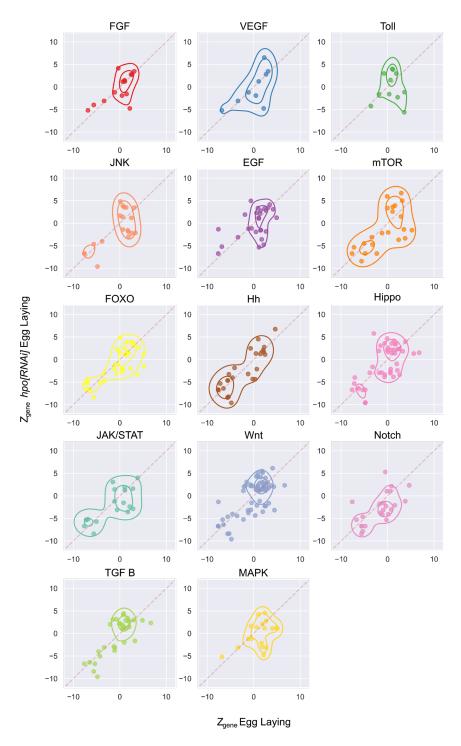


Figure S3. Comparison of egg laying candidate genes by pathway. Z_{gene} of egg laying of adult females of tj>hpo[RNAi], candidate[RNAi] plotted against Z_{gene} of egg laying of tj>candidate[RNAi] adult females displayed by pathway. Contour plots indicate a 2D gaussian kernel density estimation.

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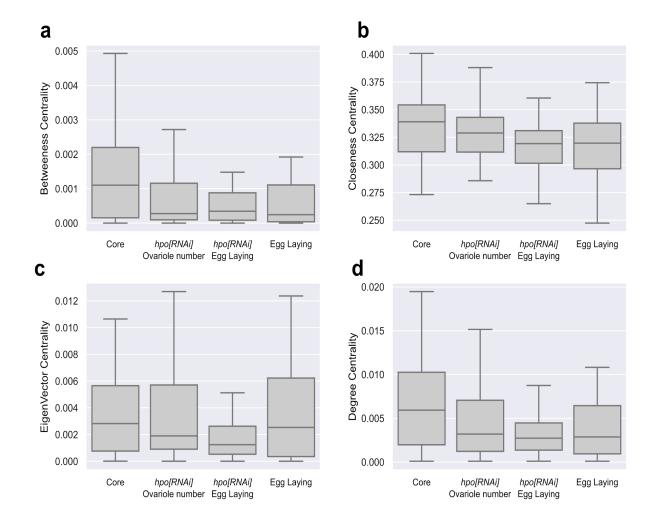


Figure S4. Box plots of the four centrality measures calculated for the genes in each of the four phenotypic modules. See modules in Figures 5 and S6.

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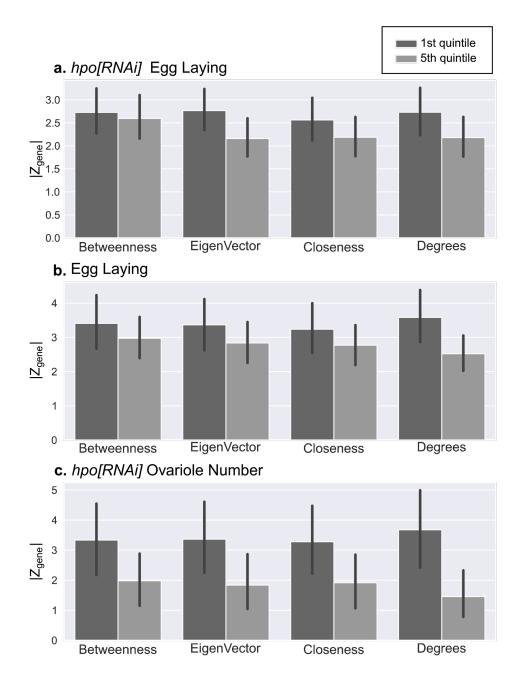
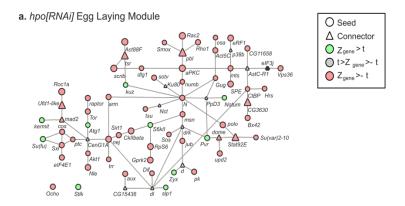
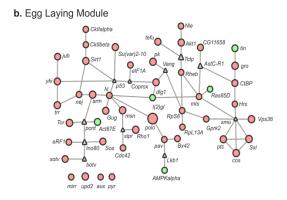


Figure S5. Comparisons of the Z_{gene} scores of the positive candidate genes sorted by centrality metrics. In each screen (a, b, c), the $|Z_{gene}|$ values of the first (dark grey) and fifth (light grey) quintiles of positive candidate genes ordered by rank for each of the four chosen centrality metrics, are plotted as a bar plot. Bars indicate standard error.





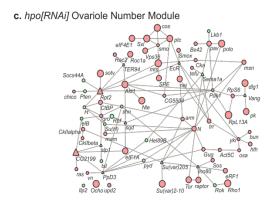


Figure S6. Three modules generated by the Seed Connector Algorithm (SCA). a) hpo[RNAi] Egg Laying Module b) Egg Laying Module c) hpo[RNAi] Ovariole Number Module. The size of the shapes indicate the Z_{gene} score of the gene. Circles = seed genes; triangles = connector genes. Green = genes with a positive Z_{gene} above the threshold. Red = genes with a negative Z_{gene} above threshold. Grey = genes with Z_{gene} values below the threshold. All connectors were phenotypically tested (Table S1) except eukaryotic translation initiation factor 3 subunit <math>j (elF3J), in the hpo[RNAi] Egg Laying Module (black triangle), for which no RNAi stock as available at the time of testing.

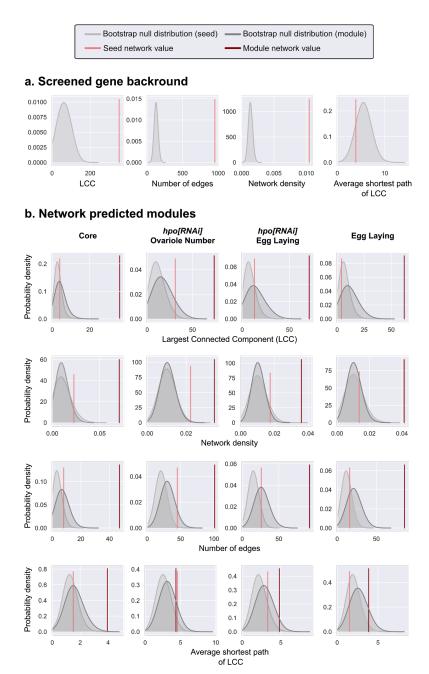


Figure S7. Comparison of network metrics before (Seed Network) and after (Module Network) application of the seed connector algorithm. a) Comparison of network metrics of all screened genes (red line) to a null distribution of network metrics derived by bootstrapping an equal number of randomly selected genes in the PPI (grey curve). b) Comparisons of the Largest Connected Component (LCC), network density, number of edges and average shortest path between the seed network (light red line) and the module network (dark red line). The bootstrapped null distribution (1000 bootstraps) of both the seed network (light grey curve) and the module network (dark grey curve) are indicated..

Supplementary tables

709 Table S1

- 710 Table S1: Tabulation of raw data and analysis for every gene in the screen.
- 711 https://github.com/extavourlab/hpo ova eggL screen/blob/master/Results/MasterTable.c
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- This table contains a summary representation of the data generated by the three screens as
- 715 well as results from the analysis. Each line corresponds to an independent measurement of a
- 716 particular RNAi line. Some genes which did not pass the first filter of $|Z_{gene}| > 1$ in the *hpo[RNAi]*
- Fig. 2 Egg Laying screen where then predicted as connectors, therefore they have two entries as they
- have been independently measured again. The Z scores have been rounded up to 4 significant
- 719 digits in this table and the Centrality metrics rounded up to 10 significant digits due to their low
- values, but the full values for both are available in the raw data files provided in the
- supplementary files in Data/Screens for the Z scores and Results for the centrality values.
- Moreover this is a summary table and does not contain values for controls as well as batch
- numbers, all are available in the supplementary files in Data/Screens.
- 725 **FbID**: FlyBase ID of the tested gene.
- 726 **CG number**: CG Number of the tested gene.
- NAME: Common name (as per FlyBase nomenclature) of the gene if existing, else it is a -.
 - **SYMBOL**: Symbol (as per FlyBase nomenclature) of the gene if existing, else CG number
- 729 [ScreenName] [Variable] (Metric) Count: Within the screen [ScreenName], the count of the
- 730 measured variable [*Variable*]. Optional: (*metric*) will indicate if a particular operation was done
- over the data, such as sum, mean or standard deviation.
- e.g. [HippoRNAi_EggL]_[Day_4_Egg]_Count is the count of eggs, on day 4, of the hpo[RNAi]
- 733 Egg Laying screen.
- [ScreenName]_[Variable]_(Metric)_Zscore: Within the screen [ScreenName], the Z score of
- the measured variable [Variable] as calculated to batch control. Optional: (*metric*) will indicate if
- a particular operation was done over the data, such as sum, mean or standard deviation.
- e.g. [EggL]_[All_Days_Egg]_(Sum)_Zscore is the Z score of the sum of eggs count, of the Egg Laying screen.
- **PPI_[Metric]_centrality**: Within the PPI used in this paper, the calculated centrality value for the metric [*Metric*].
- 741 [ModuleName] Network: Presence of absence of a gene in the module [ModuleName]. If the
- 742 gene is in the module, this value is True, if it is absent it is False. (An exception is made for the
- Meta Network displayed in Figure 7 where instead of True/False, the group assignment I-VII is written)
- [ModuleName]_Connector: Status of a gene in the module [ModuleName] as a connector. If
 True, the gene is a connector, else if False, the gene is not a connector.
- 747 [PathwayName] Pathway: Participation of a gene to the signalling pathway [PathwayName].
- If the gene participates in the pathway the value is 1, else it is 0.

749 Table S2

FbID	CG Number	Name	Symbol
FBgn0283468	CG3412	supernumerary limbs	slmb
FBgn0267821	CG5102	daughterless	da
FBgn0266724	CG5161	TRAPP subunit 20	Trs20
FBgn0267378	CG7085	sauron	sau
FBgn0267487	CG9181	Protein tyrosine phosphatase 61F	Ptp61F
FBgn0267912	CG9819	Calcineurin A at 14F	CanA-14F
FBgn0086371	CG9829	poly	poly
FBgn0267350	CG10260	Phosphatidylinositol 4- kinase III alpha	PI4KIIIalpha
FBgn0267698	CG10295	p21-activated kinase	Pak
FBgn0283462	CG18279	Immune induced molecule prepropeptide	IMPPP
FBgn0267339	CG33338	p38c MAP kinase	p38c
FBgn0085506	CG40635	-	CG40635

Table S2. 12 signalling candidate genes with no available RNAi lines at either BDSC or VDRC at the time of this study.

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Table S3

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Centrality metric	hpo[RNAi] Ovariole Number	<i>hpo[RNAi]</i> Egg Laying	Egg Laying
Betweenness	0.603	0.57	0.586
EigenVector	0.632	0.573	0.586
Closeness	0.612	0.551	0.588
Degrees	0.615	0.592	0.599

Table S3: Area under the curve (AUC) of ROC curves. AUC values for the ROC curves for each centrality measure for the three screens (Figure 4a). AUC values range from 0 to 1. A score above 0.5 indicates a positive correlation between the continuous variable (centrality) and the binary variable (above or below the Z score threshold). A score of 0.5 or less indicates no correlation between the variables.

Table S4

Module	Number of Seeds	Number of Connectors	Number of connector genes above Z _{gene} threshold within module phenotype
hpo[RNAi] Egg Laying	58	18	7 (41.2%)
Core	27	10	1 (10.0%)
Egg Laying	49	11	0 (0.0%)
<i>hpo[RNAi]</i> Ovariole Number	66	11	3 (27.3%)

Table S4. Distribution of seed genes and connectors in each module. Two genes that were above $|Z_{gene}|$ threshold (Table 2) in the hpo[RNAi] Egg Laying (CG12147) and hpo[RNAi] Ovariole Number seed list (CG6104) were not found in the PPI, and therefore not included in the network analysis or in this table (see methods for details). The removal of these two genes accounts for the difference between the number of positive candidates in Table 2 and the number of seed genes in these two modules (Table S1 and S4). The proportion of connectors whose loss of function produced a significant phenotype ($|Z_{gene}|$ above threshold) is in parentheses and plotted in Figure 6a, 6b). All connectors except eukaryotic translation initiation factor 3 subunit <math>j (elF3J) in the hpo[RNAi] Egg Laying Module, for which no RNAi line was available at the time of testing, were tested. Therefore, the percentages of connectors above the threshold for the hpo[RNAi] Egg Laying Module were calculated out of 17 connectors.

Table S5

Total number of unique connectors	Number of connector genes above $ Z_{gene} $ threshold		
in all four modules	For Egg Laying Phenotype	For <i>hpo[RNAi]</i> Egg Laying Phenotype	For <i>hpo[RNAi]</i> Ovariole Number Phenotype
43	10 (23.8%)	13 (31.0%)	14 (33.3%)

Table S5. Number of unique connector genes above $|Z_{gene}|$ threshold for the three phenotypic measurements. Percentage of the number of connectors above threshold for each phenotype from the total number of connectors is in parentheses and plotted in Figure 6c. All connectors except *eukaryotic translation initiation factor 3 subunit j* (*eIF3J*) in the *hpo[RNAi]* Egg Laying Module, for which no RNAi line was available at the time of testing, were tested. Therefore, the percentages of connectors above the threshold were calculated out of 42 unique connectors.

Methods

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LEAD CONTACT AND MATERIALS AVAILABILITY

- 789 This study did not generate new unique reagents. This study generated new python3 code
- available on GitHub: https://github.com/extavourlab/hpo ova eggL screen.
- 792 Further information and requests for resources and reagents should be directed to and will be
- 793 fulfilled by the Lead Contact, Cassandra G. Extavour (extavour@oeb.harvard.edu).

EXPERIMENTAL MODEL AND SUBJECT DETAILS

- 795 Wild type and mutant lines of *Drosophila melanogaster* were obtained from publicly accessible
- stock centers and maintained as described in "Fly Stocks" below. Genotypes and provenance
- are provided in the Key Resource Table. Candidate genes were randomly assigned to batches
- for screening (see Table S1 for which genes were in each batch). F1 animals from the same
- 799 cross were randomly assigned to experimental groups for phenotyping in all screens.

METHOD DETAILS

801 Fly stocks

- Flies were reared at 25°C at 60% humidity with standard *Drosophila* food (Sarikaya et al., 2012)
- 803 containing yeast and in uncrowded conditions as previously defined (Sarikaya and Extavour,
- 804 2015). RNAi lines were obtained from the TRiP RNAi collection at the Bloomington *Drosophila*
- 805 Stock Centre (BDSC) and from the Vienna *Drosophila* Resource Centre (VDRC). See Key
- 806 Resources Table for complete list of stocks used in this study. Oregon R was used as a wild
- type strain. The genotype of the traffic jam:Gal4 line used in the screen was y w; P{w[+mW.hs]
- 808 = GawB}NP1624 (Kyoto Stock Center, K104–055; abbreviated hereafter as tj:Gal4). The hippo
- 809 RNAi line used in the screen was y[1] v[1]; $P\{y[+t7.7]v[+t1.8]=TRiP. HMS00006\}attP2$
- 810 (BDSC:33614; abbreviated hereafter as hpo[RNAi]).

Egg and ovariole number counts

- 812 Adult egg laying was quantified by crossing three virgin females of the desired genotype (see
- "Screen design" below) with two males in a vial containing standard food and yeast granules
- 814 (day one) and then transferring them into a fresh food vial without yeast granules for a 24 hour
- period. Eggs from vials were then counted by visual inspection of the surface of the food in the
- vial. Males and females were transferred to fresh food vials without yeast granules, every day
- thereafter until day six. All egg laying measurements reported and analysed in the paper are the
- sum of the eggs laid by three adult female flies over the five days of this assay (days two

through six without yeast granules). Data from any vial in which either a female or male died, during the course of the experiment, were not included in the analysis.

Ovariole number was quantified by mating ten virgin adult females with five virgin adult Oregon R males for three days post eclosion in vials with yeast at 25°C and 60% humidity. After this three-day mating period, all 20 adult ovaries from the mated females were dissected in 1X PBS with 0.1% Triton-X-100 and stained with 1ug/ml Hoechst 33321 (1:10,000 of a 10mg/ml stock solution). Ovarioles were separated from each other with No. 5 forceps (Fine Science Tools) and counted by counting the number of germaria under a ZEISS Stemi 305 compact stereo microscope with a NIGHTSEA stereo microscope UV Fluorescence adaptor.

Screen design

In the primary screen (Figure 1a: hpo[RNAi] Egg Laying), 463 candidate genes (Table S1) were screened for the effect of an RNAi-induced loss of gene function in a hpo[RNAi] background on the number of eggs laid in the first five days of mating (see "Egg and ovariole number counts" above) by adult females. These females were the F1 offspring of UAS: candidate gene RNAi males crossed to $P\{w[+mW.hs] = GawB\}NP1624$; $P\{y[+t7.7] v[+t1.8] = TRiP.HMS00006\}$ attP2 (tj:Gal4; UAS:hpo[RNAi]) virgin adult females (Figure 1a: hpo[RNAi] Egg Laying). All genes that yielded an egg laying count with a $|Z_{qene}| > 1$ (see "Gene selection based on Z score and batch standardization" below) were selected to undergo two secondary screenings (n=273, Table 2, Figure 1d). First, these genes were screened for effects on the egg laying of mated adult female offspring from a cross of UAS:candidate gene[RNAi] males and tj:Gal4 virgin adult females (Figure 1b: Egg Laying). Secondly, these genes were screened for effects on ovariole number in a hpo[RNAi] background. All 20 ovaries from ten adult female F1 offspring of a cross between UAS:candidate gene[RNAi] males to P{w[+mW.hs] = GawB}NP1624; P{v[+t7.7] v[+t1.8]=TRiP.HMS00006}attP2 (ti:Gal4; UAS:hpo[RNAi]) virgin adult females were scored for ovariole number (see "Egg and ovariole number counts" above). (Figure 1c: hpo[RNAi] Ovariole Number).

Gene selection based on Z score and batch standardization

Candidate genes were screened in batches with an average size of 50 genes. For each batch, control flies were the female F1 offspring of Oregon R males crossed to $P\{w[+mW.hs] = GawB\}NP1624$; $P\{y[+t7.7] \ v[+t1.8] = TRiP.HMS00006\}$ attP2 (tj:Gal4; UAS:hpo[RNAi]) virgin adult females. Because the control group in each batch had slightly different distributions of egg laying and ovariole number values (Figure S1), it was inappropriate to compare absolute mean values between genes that were scored in different batches. Instead, comparisons of the Z score of each candidate (Z_{gene}) to its batch control group was used as a discriminant. This approach standardizes for batch effects and allows the comparison of all genotypes within and across the primary and secondary screens with a single metric (Z_{gene}).

Firstly, the mean and standard deviation of the eggs laid by the control genotype for a batch were calculated as μ_b and σ_b respectively. Then, using the number of eggs laid by adult females

of a candidate gene RNAi (x_{gene}) of the same batch, the Z score for the egg laying count of that gene (Z_{gene}) was calculated as $Z_{gene} = \frac{x_{geme} - \mu_b}{\sigma_b}$. The same standardization protocol was applied to both egg laying and ovariole number counts of every gene and its corresponding batch control.

Ovariole numbers were derived from counts of the number of ovarioles per ovary for 20 ovaries per candidate gene, and a threshold of $|Z_{gene}| > 2$ (corresponding to a false positive probability less than 0.045) was applied for ovariole number phenotype. Egg laying counts were derived from measurements of three females in a single vial per gene. We therefore chose to be more conservative in our Z score comparisons for the egg laying phenotype, than for ovariole number phenotype, and applied a stringent threshold of $|Z_{gene}| > 5$ (corresponding to a false positive probability less than 0.00006) to select genes of interest. All genes with $|Z_{gene}|$ values above these thresholds are referred to throughout the study as "positive candidates". (See Ipython notebooks $02_Z_score_calculation.ipynb$ and $02.2_Z_score_calculation_prediction.ipynb for code implementation and calculation of Z scores, and <math>06_Screen$ Analysis.ipynb for batch effects.)

Signaling pathway enrichment analysis

- To study the enrichment of a particular signaling pathway in a group of candidate genes that had similar phenotypic effects revealed by the screen, custom scripts (see 07_Signaling_pathway_analysis.ipynb for code implementation) were generated to implement two different methods (Figure 3a, 3b; Figure S52a-c).
- The first method is a numerical method that uses bootstrapping to calculate the null distribution of the number of members (M) of a signaling pathway (S) that would be expected at random in a set of genes of size (N). The script randomly sampled N genes from among the 463 tested *D. melanogaster* signaling genes 10,000 times, and counted the number of genes (M) that were members of the signaling pathway S. Positive candidates in each of the three screens were sorted by their presence in signalling pathways and counted. The Z score was then calculated by comparing the experimentally observed number of positive candidates in each signaling pathway against the bootstrapped null distribution.
- The second method used the hypergeometric p-value to calculate the probability of M members of a signaling pathway being in a group of N genes, given a starting population of 463 tested *D. melanogaster* signaling genes, and the known attribution to a pathway S of each gene.

Protein-Protein Interaction Network (PPI) building

There is no standard complete Protein-Protein Interaction (PPI) network available for *Drosophila*melanogaster. However, there exist many smaller networks from different screens, as well as
literature extractions. We therefore combined data from these sources and then created a PPI
for use in the present study, as follows:

Step 1: Several screens assessing protein-protein interactions have been centralized in a database called DroID: http://www.droidb.org. The version DroID_v2018_08 was used. All available datasets were first downloaded from that database using this link:

http://www.droidb.org/Downloads.jsp. The description of all of these datasets can be found here: http://www.droidb.org/DBdescription.jsp

Step 2: We used the datasets from all screens that assessed direct protein-protein interactions and did not use the interolog database (predicted protein interaction based on mouse human and yeast PPI). These direct assessment screens were seven in total, as follows:

- Finley Yeast Two-Hybrid Data (size 2.0 MB)
- Curagen Yeast Two-Hybrid Data (size 4.6 MB)
- Hybrigenics Yeast Two-Hybrid Data (size 381 KB)
- Perrimon co-AP complex (size 108 KB)
- DPiM co-AP complex (size 6.3 MB)
- PPI from other databases (size 16.2 MB)
- PPI curated by FlyBase (size 7.4 MB)

An important element to note is that the PPI curated by FlyBase is a literature-based PPI. FlyBase protein-protein interactions are experimentally derived physical interactions curated from the literature by FlyBase, and does not include FlyBase-curated genetic interactions.

Step 3: We concatenated the seven datasets listed above into a single unique database. A custom python script was created that downloads and reads each of the above seven unique PPI tables, and generates a single PPI network. From this concatenation, a single edge undirected network was created and saved. This network is hereafter referred to as **the PPI** (see 01 PPI builder.ipynb).

Network metric computations

The centrality of a node is often used as a measure of a node's importance in a network. Within a PPI, the centrality of a gene reflects the number of interactions in which the gene directly or indirectly participates. Four different centrality metrics were computed for all genes in the PPI using the networkx python library:

- (1) **Betweenness** reflects the number of shortest paths passing through a gene.
- (2) **Eigenvector** is a measure of the influence of a gene in the network.
- (3) **Closeness** measures the sum of shortest distance of a gene to all the other genes.
- (4) **Degree centrality** corresponds to the normalized number of edges of a gene in the network.

While there exist more centrality measures, these four are commonly used to assess biological networks. These computed centrality parameters of the genes measured in the screen were

computed with 03_ROC_curve_analysis_of_network_metrics.ipynb, and are reported in the Table S1 (see 09 Making the database table.ipynb).

Receiver Operating Characteristic (ROC) curves

To check whether the centrality of a gene in the network could predict the phenotypic effect produced by RNAi against that gene, ROC curves were plotted for the four aforementioned centrality measures of each gene in each screen. A ROC analysis is used to measure the correlation between a continuous variable (centrality) and a binary outcome (above or below Z score threshold). Therefore, for each screen, measured genes were rank-ordered from high centrality to low centrality, and plotted against the binary outcome of $|Z_{gene}|$ being above or below the appropriate |Z| score threshold (>5 for egg laying and >2 for ovariole number). The Area Under the Curve (AUC) measures the extent of correlation between centrality and effect of a gene on measured phenotype. AUC above or below 0.5 indicates a positive or negative correlation respectively, while an AUC of 0.5 indicates no correlation of the parameters. The scikit learn python package was used to calculate the AUC of each ROC curve plotted (see 03 ROC curve analysis of network metrics.ipynb).

Building the network modules

Network modules were built using the previously published Seed-Connector algorithm (SCA) (Wang et al., 2017; Wang and Loscalzo, 2018), implemented here in python (see 04 Seed-Connector.ipynb) and illustrated in Figure 5a. Creating a module using the SCA requires a list of seed genes and a PPI. From each of the three screens, we selected the genes whose $|Z_{vene}|$ value was above the threshold and created three seed lists respectively (Figure 4c: Egg laying, hpo[RNAi] egg laying and hpo[RNAi] ovariole 'seed' list). A fourth list consisting of the intersection of the aforementioned seed lists was also collated and called the core 'seed' list (Figure 4b). Genes were assigned in the core list if they passed the Z threshold in all 3 screens. The Seed-Connector algorithm was then executed on each of these seed lists using the PPI. Not all genes in the four seed lists were found in the PPI network (specifically, CG12147 in the hpo[RNAi] Egg Laving seed list and CG6104 in the hpo[RNAi] Ovariole number seed list were absent from the PPI) and were therefore eliminated from further network analysis. The removal of these two genes accounts for the variation in the number of positive candidates in Table 2 and the number of seed genes in the module. Modules were obtained for each seed list (Figure 5b; Figure S6) consisting of the seed genes (circles in Figure 5b and Figure S6) and previously untested genes added by the SCA (squares in Figure 5b and Figure S6) to increase the LCC size that we refer to as connector genes (see 04 Seed-Connector.ipynb). The results of the algorithm are summarized in Table S1.

The modularity of the subnetworks was then assessed using four network metrics namely Largest Connected Component (LCC), number of edges, network density and average shortest path in the LCC. Each metric for each module was assessed using distance of the network metric to a null distribution. Initially, the null distribution was calculated by taking 1000 samples of 463 genes randomly selected from the PPI and calculating the above metrics. We found that

the 463 genes selected in the signalling screen were already more connected than the null 979 980 distribution of sets of 463 genes randomly selected from the PPI (Figure S7a). Therefore, to 981 avoid a false positive detection of modularity, the four experimentally obtained subnetworks 982 were compared to null distributions obtained by randomly sampling an equal number of genes 983 from the 463 signalling candidate genes selected for our screen. For each of the four modules, 984 comparison of the metrics was performed on the seed lists and the sub-network after the SCA. 985 Most metrics were enriched in the seed group when compared to the null distribution with the 986 exception of the Average shortest path (Figure S7b, light red line). The sub-networks obtained 987 from the SCA further increased all four metrics suggesting the modularity of the four sub-988 networks (Figure S7b, dark red line; see 05 Network Module testing.ipynb for code 989 implementation).

Meta network

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To build the meta network, the genes from all four modules were concatenated into one network. The network was then visually sorted in an approach akin to projecting the network on a Venn Diagram. The meta network was sorted by which of the three screens the gene was positive in. The intersections were genes whose $|Z_{gene}|$ value was above the threshold in more than one and possibly all three of the screening paradigms. For example, if a gene was found in the hpo[RNAi] Ovariole Number and Egg Laying module it is then assigned to the dual positive group hpo[RNAi] Ovariole Number / Egg Laying (Figure 7a, module VI). After applying this grouping strategy, the connectivity across the groups was studied by calculating the edge density between all groups $(density = \frac{Edges_{1,2}}{Nodes_1*Nodes_2})$. Finally, the proportion of each signaling candidate in each of those groups was calculated by taking the number of members of a signaling pathway divided by the total members of a group (see Ipython notebook 08 MetaModule Analysis.ipynb).

QUANTIFICATION AND STATISTICAL ANALYSIS

Number of samples

1005 The number of samples across the different screens were as follows:

hpo[RNAi] Egg Laying and Egg Laying screens

- Controls: five vials of three females and two males
- 1008 Sample: one vial of three females and two males

hpo[RNAi] Ovariole number screen

- Controls: 20 flies, two ovaries per fly considered as independent measurements
- 1011 Sample: 10 flies, two ovaries per fly considered as independent measurements

Correction of batch effect

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- Despite best efforts to maintain the exact same condition between each experiment, some
- 1014 variation was measured between the batches. Control flies showed variations in both measured
- 1015 phenotypes, ovariole number and egg laying (Figure S1). In order to compare the values
- 1016 measured across different batches, each sample was standardized by calculating its Z score
- 1017 (Z_{gene}) to the control distribution. For each batch, the measurements for controls were pooled into
- 1018 a distribution, and the mean and standard deviation was computed. Then each sample was
- 1019 compared to its respective batch and its Z score computed (see "Gene selection based on Z
- 1020 score and batch standardization" for formula).

Statistical analysis

- All statistical analyses were performed using the scipy stats module (https://www.scipy.org/) and
- scikit learn (https://scikit-learn.org/). Significance thresholds for p-values were set at 0.05.
- Statistical tests and p-values are reported in the figure legends. All statistical tests can be found
- in the lpython notebooks mentioned below.

DATA AND CODE AVAILABILITY

- This study generated a series of python3 lpython notebook files that perform the entire analysis
- presented in this study. All the results presented in this paper, including the figures with the
- 1029 exception of the network visualizations, which were created using Cytoscape3
- 1030 (https://cytoscape.org/) can be reproduced by running the aforementioned python3 code. The
- 1031 raw data, calculations made with these data, and code used for calculations and analyses
- 1032 (Ipython notebooks) are available as supplementary information. For ease of access, legibility
- and reproducibility, the code and datasets have been deposited in a GitHub repository available
- 1034 at https://github.com/extavourlab/hpo_ova_eggL_screen.

KEY RESOURCES TABLES

Software and libraries

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All software and libraries used in this study are published under open source licenses and are therefore publicly available.

Туре	Name	Version	Source
Library	matplotlib	3.0.0	https://matplotlib.org/
Library	networkx	2.3	http://networkx.github.io/
Library	numpy	1.11.3	https://www.numpy.org/
Library	pandas	0.20.3	https://pandas.pydata.org /
Library	progressbar	3.38.0	https://github.com/niltonvolpato/python-progressbar
Library	scipy	1.1.0	https://www.scipy.org/
Library	seaborn	0.9.0	https://seaborn.pydata.org/
Software	Cytoscape	3.4.0	https://cytoscape.org/
Software	Inkscape	0.92.3	https://inkscape.org/
Software	Python3	3.7	https://www.python.org/

Drosophila melanogaster genetic lines

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Experimental Models: Organisms/Strains			
Description	Stock Center	IDs	
D. melanogaster. Expresses dsRNA for RNAi of CanA1 (FBgn0010015) under UAS control in the VALIUM10 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.JF01871}attP2	Bloomington Drosophila Stock Center	BDSC:25850; FlyBase:FBst0025850 ;	
D. melanogaster. Expresses dsRNA for RNAi of fng (FBgn0011591) under UAS control in the VALIUM10 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.JF01967}attP2	Bloomington Drosophila Stock Center	BDSC:25947; FlyBase:FBst0025947	
D. melanogaster. Expresses dsRNA for RNAi of Aplip1 (FBgn0040281) under UAS control in the VALIUM10 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.JF02049}attP2	Bloomington Drosophila Stock Center	BDSC:26024; FlyBase:FBst0026024 ;	
D. melanogaster. Expresses dsRNA for RNAi of E(spl)mdelta-HLH (FBgn0002734) under UAS control in the VALIUM10 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.JF02101}attP2/TM3, Sb[1]	Bloomington Drosophila Stock Center	BDSC:26203; FlyBase:FBst0026203 ;	
D. melanogaster. Expresses dsRNA for RNAi of sima (FBgn0266411) under UAS control in the VALIUM10 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.JF02105}attP2	Bloomington Drosophila Stock Center	BDSC:26207; FlyBase:FBst0026207;	
D. melanogaster. Expresses dsRNA for RNAi of E(spl)m8-HLH (FBgn0000591) under UAS control in the VALIUM10 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.JF02096}attP2	Bloomington Drosophila Stock Center	BDSC:26322; FlyBase:FBst0026322 ;	
D. melanogaster. Expresses dsRNA for RNAi of pan (FBgn0085432) under UAS control in the VALIUM10 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.JF02306}attP2	Bloomington Drosophila Stock Center	BDSC:26743; FlyBase:FBst0026743	
D. melanogaster. Expresses dsRNA for RNAi of CanB (FBgn0010014) under UAS control in the VALIUM10 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.JF02616}attP2	Bloomington Drosophila Stock Center	BDSC:27307; FlyBase:FBst0027307	
D. melanogaster. Expresses dsRNA for RNAi of mib1 (FBgn0263601) under UAS control in the VALIUM10 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.JF02629}attP2	Bloomington Drosophila Stock Center	BDSC:27320; FlyBase:FBst0027320 ;	
D. melanogaster. Expresses dsRNA for RNAi of Nct (FBgn0039234) under UAS control in the VALIUM10	Bloomington Drosophila Stock Center	BDSC:27498; FlyBase:FBst0027498 ;	

vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.JF02648}attP2		
D. melanogaster. Expresses dsRNA for RNAi of Cbl (FBgn0020224) under UAS control in the VALIUM10 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.JF02650}attP2	Bloomington Drosophila Stock Center	BDSC:27500; FlyBase:FBst0027500
D. melanogaster. Expresses dsRNA for RNAi of Atg12 (FBgn0036255) under UAS control in the VALIUM10 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.JF02704}attP2	Bloomington Drosophila Stock Center	BDSC:27552; FlyBase:FBst0027552 ;
D. melanogaster. Expresses dsRNA for RNAi of fz2 (FBgn0016797) under UAS control in the VALIUM10 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.JF02722}attP2	Bloomington Drosophila Stock Center	BDSC:27568; FlyBase:FBst0027568 ;
D. melanogaster. Expresses dsRNA for RNAi of Psn (FBgn0284421) under UAS control in the VALIUM10 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.JF02760}attP2/TM3, Sb[1]	Bloomington Drosophila Stock Center	BDSC:27681; FlyBase:FBst0027681 ;
D. melanogaster. Expresses dsRNA for RNAi of Pi3K92E (FBgn0015279) under UAS control in the VALIUM10 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.JF02770}attP2/TM3, Sb[1]	Bloomington Drosophila Stock Center	BDSC:27690; FlyBase:FBst0027690 ;
D. melanogaster. Expresses dsRNA for RNAi of Cdk4 (FBgn0016131) under UAS control in the VALIUM10 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.JF02795}attP2	Bloomington Drosophila Stock Center	BDSC:27714; FlyBase:FBst0027714 ;
D. melanogaster. Expresses dsRNA for RNAi of mts (FBgn0004177) under UAS control in the VALIUM10 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.JF02805}attP2	Bloomington Drosophila Stock Center	BDSC:27723; FlyBase:FBst0027723 ;
D. melanogaster. Expresses dsRNA for RNAi of Pdk1 (FBgn0020386) under UAS control in the VALIUM10 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.JF02807}attP2	Bloomington Drosophila Stock Center	BDSC:27725; FlyBase:FBst0027725 ;
D. melanogaster. Expresses dsRNA for RNAi of ds (FBgn0284247) under UAS control in the VALIUM10 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.JF02842}attP2	Bloomington Drosophila Stock Center	BDSC:28008; FlyBase:FBst0028008 ;
D. melanogaster. Expresses dsRNA for RNAi of Hrs (FBgn0031450) under UAS control in the VALIUM10 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.JF02860}attP2	Bloomington Drosophila Stock Center	BDSC:28026; FlyBase:FBst0028026 ;

D. melanogaster. Expresses dsRNA for RNAi of mam (FBgn0002643) under UAS control in the VALIUM10 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.JF02881}attP2	Bloomington Drosophila Stock Center	BDSC:28046; FlyBase:FBst0028046 ;
D. melanogaster. Expresses dsRNA for RNAi of bun (FBgn0259176) under UAS control in the VALIUM10 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.JF02954}attP2	Bloomington Drosophila Stock Center	BDSC:28322; FlyBase:FBst0028322 ;
D. melanogaster. Expresses dsRNA for RNAi of aos (FBgn0004569) under UAS control in the VALIUM10 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.JF03020}attP2	Bloomington Drosophila Stock Center	BDSC:28383; FlyBase:FBst0028383 ;
D. melanogaster. Expresses dsRNA for RNAi of pcx (FBgn0003048) under UAS control in the VALIUM10 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.HM05038}attP2/TM3, Sb[1]	Bloomington Drosophila Stock Center	BDSC:28552; FlyBase:FBst0028552 ;
D. melanogaster. Expresses dsRNA for RNAi of Su(fu) (FBgn0005355) under UAS control in the VALIUM10 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.HM05045}attP2	Bloomington Drosophila Stock Center	BDSC:28559; FlyBase:FBst0028559 ;
D. melanogaster. Expresses dsRNA for RNAi of Apc2 (FBgn0026598) under UAS control in the VALIUM10 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.HM05073}attP2	Bloomington Drosophila Stock Center	BDSC:28585; FlyBase:FBst0028585 ;
D. melanogaster. Expresses dsRNA for RNAi of I(2)tid (FBgn0002174) under UAS control in the VALIUM10 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.HM05082}attP2	Bloomington Drosophila Stock Center	BDSC:28594; FlyBase:FBst0028594 ;
D. melanogaster. Expresses dsRNA for RNAi of Su(H) (FBgn0004837) under UAS control in the VALIUM10 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.HM05110}attP2/TM3, Sb[1]	Bloomington Drosophila Stock Center	BDSC:28900; FlyBase:FBst0028900 ;
D. melanogaster. Expresses dsRNA for RNAi of PDZ-GEF (FBgn0265778) under UAS control in the VALIUM10 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.HM05139}attP2	Bloomington Drosophila Stock Center	BDSC:28928; FlyBase:FBst0028928 ;
D. melanogaster. Expresses dsRNA for RNAi of wntD (FBgn0038134) under UAS control in the VALIUM10 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.HM05158}attP2	Bloomington Drosophila Stock Center	BDSC:28947; FlyBase:FBst0028947
D. melanogaster. Expresses dsRNA for RNAi of Cdk2 (FBgn0004107) under UAS control in the VALIUM10 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.HM05163}attP2	Bloomington Drosophila Stock Center	BDSC:28952; FlyBase:FBst0028952 ;

Bloomington	BDSC:28984;
Drosophila Stock	FlyBase:FBst0028984
Center	;
Bloomington	BDSC:29324;
Drosophila Stock	FlyBase:FBst0029324
Center	;
Bloomington	BDSC:29378;
Drosophila Stock	FlyBase:FBst0029378
Center	;
Bloomington	BDSC:29442;
Drosophila Stock	FlyBase:FBst0029442
Center	;
Bloomington	BDSC:30513;
Drosophila Stock	FlyBase:FBst0030513
Center	;
Bloomington	BDSC:31037;
Drosophila Stock	FlyBase:FBst0031037
Center	;
Bloomington	BDSC:31067;
Drosophila Stock	FlyBase:FBst0031067
Center	;
Bloomington	BDSC:31183;
Drosophila Stock	FlyBase:FBst0031183
Center	;
Bloomington Drosophila Stock Center	BDSC:31197; FlyBase:FBst0031197
Bloomington	BDSC:31228;
Drosophila Stock	FlyBase:FBst0031228
Center	;
Bloomington	BDSC:31307;
Drosophila Stock	FlyBase:FBst0031307
Center	;
	Drosophila Stock Center Bloomington Drosophila Stock Center

D. melanogaster. Expresses dsRNA for RNAi of TI (FBgn0262473) under UAS control in the VALIUM1 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.JF01276}attP2	Bloomington Drosophila Stock Center	BDSC:31477; FlyBase:FBst0031477;
D. melanogaster. Expresses dsRNA for RNAi of I(2)gl (FBgn0002121) under UAS control in the VALIUM1 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.JF01073}attP2	Bloomington Drosophila Stock Center	BDSC:31517; FlyBase:FBst0031517 ;
D. melanogaster. Expresses dsRNA for RNAi of Jra (FBgn0001291) under UAS control in the VALIUM1 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.JF01184}attP2	Bloomington Drosophila Stock Center	BDSC:31595; FlyBase:FBst0031595 ;
D. melanogaster. Expresses dsRNA for RNAi of Axn (FBgn0026597) under UAS control in the VALIUM1 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.HM04012}attP2	Bloomington Drosophila Stock Center	BDSC:31705; FlyBase:FBst0031705 ;
D. melanogaster. Expresses dsRNA for RNAi of gig (FBgn0005198) under UAS control in the VALIUM1 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.HM04083}attP2	Bloomington Drosophila Stock Center	BDSC:31770; FlyBase:FBst0031770 ;
D. melanogaster. Expresses dsRNA for RNAi of Med (FBgn0011655) under UAS control in the VALIUM10 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.JF02218}attP2	Bloomington Drosophila Stock Center	BDSC:31928; FlyBase:FBst0031928 ;
D. melanogaster. Expresses dsRNA for RNAi of RagC-D (FBgn0033272) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00333}attP2	Bloomington Drosophila Stock Center	BDSC:32342; FlyBase:FBst0032342 ;
D. melanogaster. Expresses dsRNA for RNAi of AMPKalpha (FBgn0023169) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00362}attP2	Bloomington Drosophila Stock Center	BDSC:32371; FlyBase:FBst0032371 ;
D. melanogaster. Expresses dsRNA for RNAi of Rho1 (FBgn0014020) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00375}attP2/TM3, Sb[1]	Bloomington Drosophila Stock Center	BDSC:32383; FlyBase:FBst0032383 ;
D. melanogaster. Expresses dsRNA for RNAi of pk (FBgn0003090) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00408}attP2	Bloomington Drosophila Stock Center	BDSC:32413; FlyBase:FBst0032413 ;
D. melanogaster. Expresses dsRNA for RNAi of RpS6 (FBgn0261592) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00413}attP2	Bloomington Drosophila Stock Center	BDSC:32418; FlyBase:FBst0032418 ;

Bloomington Drosophila Stock Center	BDSC:32427; FlyBase:FBst0032427;
Bloomington	BDSC:32438;
Drosophila Stock	FlyBase:FBst0032438
Center	;
Bloomington	BDSC:32475;
Drosophila Stock	FlyBase:FBst0032475
Center	;
Bloomington	BDSC:32489;
Drosophila Stock	FlyBase:FBst0032489
Center	;
Bloomington	BDSC:32852;
Drosophila Stock	FlyBase:FBst0032852
Center	;
Bloomington Drosophila Stock Center	BDSC:32859; FlyBase:FBst0032859
Bloomington	BDSC:32861;
Drosophila Stock	FlyBase:FBst0032861
Center	;
Bloomington	BDSC:32862;
Drosophila Stock	FlyBase:FBst0032862
Center	;
Bloomington	BDSC:32870;
Drosophila Stock	FlyBase:FBst0032870
Center	;
Bloomington Drosophila Stock Center	BDSC:32875; FlyBase:FBst0032875;
Bloomington	BDSC:32889;
Drosophila Stock	FlyBase:FBst0032889
Center	;
	Drosophila Stock Center Bloomington Drosophila Stock Center

<u></u>	1	1
D. melanogaster. Expresses dsRNA for RNAi of Hel89B (FBgn0022787) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00684}attP2	Bloomington Drosophila Stock Center	BDSC:32895; FlyBase:FBst0032895 ;
D. melanogaster. Expresses dsRNA for RNAi of Tctp (FBgn0037874) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00701}attP2/TM3, Sb[1]	Bloomington Drosophila Stock Center	BDSC:32911; FlyBase:FBst0032911 ;
D. melanogaster. Expresses dsRNA for RNAi of jub (FBgn0030530) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00714}attP2	Bloomington Drosophila Stock Center	BDSC:32923; FlyBase:FBst0032923 ;
D. melanogaster. Expresses dsRNA for RNAi of Gug (FBgn0010825) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00756}attP2	Bloomington Drosophila Stock Center	BDSC:32961; FlyBase:FBst0032961 ;
D. melanogaster. Expresses dsRNA for RNAi of sav (FBgn0053193) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00760}attP2	Bloomington Drosophila Stock Center	BDSC:32965; FlyBase:FBst0032965;
D. melanogaster. Expresses dsRNA for RNAi of bsk (FBgn0000229) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00777}attP2	Bloomington Drosophila Stock Center	BDSC:32977; FlyBase:FBst0032977
D. melanogaster. Expresses dsRNA for RNAi of Pgcl (FBgn0011822) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00792}attP2/TM3, Sb[1]	Bloomington Drosophila Stock Center	BDSC:32992; FlyBase:FBst0032992 ;
D. melanogaster. Expresses dsRNA for RNAi of polo (FBgn0003124) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00530}attP2	Bloomington Drosophila Stock Center	BDSC:33042; FlyBase:FBst0033042 ;
D. melanogaster. Expresses dsRNA for RNAi of cnk (FBgn0286070) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00238}attP2	Bloomington Drosophila Stock Center	BDSC:33366; FlyBase:FBst0033366 ;
D. melanogaster. Expresses dsRNA for RNAi of kay (FBgn0001297) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00254}attP2	Bloomington Drosophila Stock Center	BDSC:33379; FlyBase:FBst0033379 ;
D. melanogaster. Expresses dsRNA for RNAi of apolpp (FBgn0087002) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00265}attP2/TM3, Sb[1]	Bloomington Drosophila Stock Center	BDSC:33388; FlyBase:FBst0033388 ;

V +11.8 =TRIP.HMS00278 attP2			
(FBgn002s323) under UAS control in the VALIUM20 vector: y[1] set[1] y[1] sev[2]; P[y[+17.7] brosophila Stock Center	Su(var)205 (FBgn0003607) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7]	Drosophila Stock	BDSC:33400; FlyBase:FBst0033400 ;
FBgn0261456 under UAS control in the VALIUM20 Drosophila Stock FlyBase:FBst003361 V[+t1.8]=TRIP.HMS00006]attP2 D. melanogaster. Expresses dsRNA for RNAi of Akt1 FlyBase:FBst003361 PlyBase:FBst003361 FlyBase:FBst003361 PlyBase:FBst003361 PlyBase:FBst003362 PlyBase:FBst003362 PlyBase:FBst003362 PlyBase:FBst003362 PlyBase:FBst003363 Ply	(FBgn0026323) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7]	Drosophila Stock	BDSC:33404; FlyBase:FBst0033404 ;
(FBgn0010379) under UAS control in the VALIUM20 Bloomington Drosophila Stock Center V[+11.8]=TRIP.HMS00007}attP2 Center Cent	(FBgn0261456) under UAS control in the VALIUM20 vector.: y[1] v[1]; P{y[+t7.7]	Drosophila Stock	BDSC:33614; FlyBase:FBst0033614 ;
(FBgn0004647) under UAS control in the VALIUM20 Bloomington Drosophila Stock FlyBase:FBst003361 V[+t1.8]=TRiP.HMS00009}sttP2 Center ;	(FBgn0010379) under UAS control in the VALIUM20 vector.: y[1] v[1]; P{y[+t7.7]	Drosophila Stock	BDSC:33615; FlyBase:FBst0033615 ;
(FBgn000382) under UAS control in the VALIUM20 vector.: y[1] v[1]; P{y[+t7.7] vector.: y[1] v[1]; P{y[-t7.7] vector.: y[1] v	(FBgn0004647) under UAS control in the VALIUM20 vector.: y[1] v[1]; P{y[+t7.7]	Drosophila Stock	BDSC:33616; FlyBase:FBst0033616 ;
(FBgn0003733) under UAS control in the VALIUM20 vector.: y[1] v[1]; P{y[+t7.7] vector.: y[1] v[1]; P{y[-t7.7] vector.: y[1]	(FBgn0000382) under UAS control in the VALIUM20 vector.: y[1] v[1]; P{y[+t7.7]	Drosophila Stock	BDSC:33619; FlyBase:FBst0033619 ;
Stat92E (FBgn0016917) under UAS control in the VALIUM20 vector.: y[1] v[1]; P{y[+t7.7]} Drosophila Stock v[+t1.8]=TRiP.HMS00035}attP2 Center ; D. melanogaster. Expresses dsRNA for RNAi of Dsor1 (FBgn0010269) under UAS control in the VALIUM20 vector.: y[1] v[1]; P{y[+t7.7]} Drosophila Stock v[+t1.8]=TRiP.HMS00037}attP2 Center ; D. melanogaster. Expresses dsRNA for RNAi of Pten (FBgn0026379) under UAS control in the VALIUM20 vector.: y[1] v[1]; P{y[+t7.7]} Drosophila Stock vector.: y[1] v[1]; P{y[-t7.7]} Drosophila Stock	(FBgn0003733) under UAS control in the VALIUM20 vector.: y[1] v[1]; P{y[+t7.7]	Drosophila Stock	BDSC:33627; FlyBase:FBst0033627 ;
(FBgn0010269) under UAS control in the VALIUM20 vector.: y[1] v[1]; P{y[+t7.7] Drosophila Stock v[+t1.8]=TRiP.HMS00037}attP2 Center ; D. melanogaster. Expresses dsRNA for RNAi of Pten (FBgn0026379) under UAS control in the VALIUM20 Bloomington vector.: y[1] v[1]; P{y[+t7.7] Drosophila Stock v[+t1.8]=TRiP.HMS00044}attP2 Center ; D. melanogaster. Expresses dsRNA for RNAi of CycD (FBgn0010315) under UAS control in the VALIUM20 Bloomington BDSC:33643; FlyBase:FBst003364 Center ;	Stat92E (FBgn0016917) under UAS control in the VALIUM20 vector.: y[1] v[1]; P{y[+t7.7]	Drosophila Stock	BDSC:33637; FlyBase:FBst0033637 ;
(FBgn0026379) under UAS control in the VALIUM20 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00044}attP2 D. melanogaster. Expresses dsRNA for RNAi of CycD (FBgn0010315) under UAS control in the VALIUM20 Bloomington Drosophila Stock Center Center BDSC:33643; FlyBase:FBst003364 ; BDSC:33653;	(FBgn0010269) under UAS control in the VALIUM20 vector.: y[1] v[1]; P{y[+t7.7]	Drosophila Stock	BDSC:33639; FlyBase:FBst0033639 ;
(FBgn0010315) under UAS control in the VALIUM20 Bloomington BDSC:33653;	(FBgn0026379) under UAS control in the VALIUM20 vector.: y[1] v[1]; P{y[+t7.7]	Drosophila Stock	BDSC:33643; FlyBase:FBst0033643 ;
vector y[1] sc[] v[1] sev[21], P{y[+t7.7] Drosoprilla Stock PlyBase.PBst003365 v[+t1.8]=TRiP.HMS00059}attP2 Center ;	(FBgn0010315) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7]	Drosophila Stock	BDSC:33653; FlyBase:FBst0033653 ;

Bloomington	BDSC:33661;
Drosophila Stock	FlyBase:FBst0033661
Center	;
Bloomington	BDSC:33680;
Drosophila Stock	FlyBase:FBst0033680
Center	;
Bloomington	BDSC:33681;
Drosophila Stock	FlyBase:FBst0033681
Center	;
Bloomington	BDSC:33682;
Drosophila Stock	FlyBase:FBst0033682
Center	;
Bloomington	BDSC:33683;
Drosophila Stock	FlyBase:FBst0033683
Center	;
Bloomington	BDSC:33684;
Drosophila Stock	FlyBase:FBst0033684
Center	;
Bloomington	BDSC:33754;
Drosophila Stock	FlyBase:FBst0033754
Center	;
Bloomington	BDSC:33759;
Drosophila Stock	FlyBase:FBst0033759
Center	;
Bloomington	BDSC:33768;
Drosophila Stock	FlyBase:FBst0033768
Center	;
Bloomington	BDSC:33902;
Drosophila Stock	FlyBase:FBst0033902
Center	;
Bloomington	BDSC:33915;
Drosophila Stock	FlyBase:FBst0033915
Center	;
	Drosophila Stock Center Bloomington Drosophila Stock Center

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D. melanogaster. Expresses dsRNA for RNAi of SkpC (FBgn0026175) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00871}attP2	Bloomington Drosophila Stock Center	BDSC:33925; FlyBase:FBst0033925 ;
D. melanogaster. Expresses dsRNA for RNAi of SPE (FBgn0039102) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00873}attP2	Bloomington Drosophila Stock Center	BDSC:33926; FlyBase:FBst0033926 ;
D. melanogaster. Expresses dsRNA for RNAi of Traf6 (FBgn0265464) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00880}attP2	Bloomington Drosophila Stock Center	BDSC:33931; FlyBase:FBst0033931 ;
D. melanogaster. Expresses dsRNA for RNAi of upd2 (FBgn0030904) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00901}attP2	Bloomington Drosophila Stock Center	BDSC:33949; FlyBase:FBst0033949 ;
D. melanogaster. Expresses dsRNA for RNAi of Tor (FBgn0021796) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00904}attP2	Bloomington Drosophila Stock Center	BDSC:33951; FlyBase:FBst0033951 ;
D. melanogaster. Expresses dsRNA for RNAi of dally (FBgn0263930) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00905}attP2	Bloomington Drosophila Stock Center	BDSC:33952; FlyBase:FBst0033952 ;
D. melanogaster. Expresses dsRNA for RNAi of Rheb (FBgn0041191) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00923}attP2	Bloomington Drosophila Stock Center	BDSC:33966; FlyBase:FBst0033966 ;
D. melanogaster. Expresses dsRNA for RNAi of Cat (FBgn0000261) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00990}attP2	Bloomington Drosophila Stock Center	BDSC:34020; FlyBase:FBst0034020 ;
D. melanogaster. Expresses dsRNA for RNAi of CycK (FBgn0025674) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01003}attP2	Bloomington Drosophila Stock Center	BDSC:34032; FlyBase:FBst0034032 ;
D. melanogaster. Expresses dsRNA for RNAi of wts (FBgn0011739) under UAS control in the VALIUM20 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00026}attP2	Bloomington Drosophila Stock Center	BDSC:34064; FlyBase:FBst0034064 ;
D. melanogaster. Expresses dsRNA for RNAi of yki (FBgn0034970) under UAS control in the VALIUM20 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00041}attP2	Bloomington Drosophila Stock Center	BDSC:34067; FlyBase:FBst0034067

Bloomington Drosophila Stock Center	BDSC:34080; FlyBase:FBst0034080
Bloomington	BDSC:34084;
Drosophila Stock	FlyBase:FBst0034084
Center	;
Bloomington	BDSC:34096;
Drosophila Stock	FlyBase:FBst0034096
Center	;
Bloomington	BDSC:34320;
Drosophila Stock	FlyBase:FBst0034320
Center	;
Bloomington	BDSC:34321;
Drosophila Stock	FlyBase:FBst0034321
Center	;
Bloomington	BDSC:34323;
Drosophila Stock	FlyBase:FBst0034323
Center	;
Bloomington	BDSC:34325;
Drosophila Stock	FlyBase:FBst0034325
Center	;
Bloomington	BDSC:34329;
Drosophila Stock	FlyBase:FBst0034329
Center	;
Bloomington	BDSC:34332;
Drosophila Stock	FlyBase:FBst0034332
Center	;
Bloomington	BDSC:34354;
Drosophila Stock	FlyBase:FBst0034354
Center	;
Bloomington	BDSC:34361;
Drosophila Stock	FlyBase:FBst0034361
Center	;
	Drosophila Stock Center Bloomington Drosophila Stock Center

Bloomington	BDSC:34362;
Drosophila Stock	FlyBase:FBst0034362
Center	;
Bloomington	BDSC:34392;
Drosophila Stock	FlyBase:FBst0034392
Center	;
Bloomington	BDSC:34393;
Drosophila Stock	FlyBase:FBst0034393
Center	;
Bloomington	BDSC:34522;
Drosophila Stock	FlyBase:FBst0034522
Center	;
Bloomington	BDSC:34572;
Drosophila Stock	FlyBase:FBst0034572
Center	;
Bloomington	BDSC:34590;
Drosophila Stock	FlyBase:FBst0034590
Center	;
Bloomington Drosophila Stock Center	BDSC:34603; FlyBase:FBst0034603
Bloomington	BDSC:34613;
Drosophila Stock	FlyBase:FBst0034613
Center	;
Bloomington	BDSC:34618;
Drosophila Stock	FlyBase:FBst0034618
Center	;
Bloomington	BDSC:34620;
Drosophila Stock	FlyBase:FBst0034620
Center	;
Bloomington	BDSC:34633;
Drosophila Stock	FlyBase:FBst0034633
Center	;
	Drosophila Stock Center Bloomington Drosophila Stock Center

Bloomington Drosophila Stock Center	BDSC:34634; FlyBase:FBst0034634 ;
Bloomington Drosophila Stock Center	BDSC:34637; FlyBase:FBst0034637;
Bloomington Drosophila Stock Center	BDSC:34643; FlyBase:FBst0034643 ;
Bloomington Drosophila Stock Center	BDSC:34644; FlyBase:FBst0034644 ;
Bloomington Drosophila Stock Center	BDSC:34645; FlyBase:FBst0034645 ;
Bloomington Drosophila Stock Center	BDSC:34653; FlyBase:FBst0034653
Bloomington Drosophila Stock Center	BDSC:34689; FlyBase:FBst0034689
Bloomington Drosophila Stock Center	BDSC:34700; FlyBase:FBst0034700
Bloomington Drosophila Stock Center	BDSC:34703; FlyBase:FBst0034703
Bloomington Drosophila Stock Center	BDSC:34706; FlyBase:FBst0034706 ;
Bloomington Drosophila Stock Center	BDSC:34716; FlyBase:FBst0034716 ;
	Drosophila Stock Center Bloomington Drosophila Stock Center

Bloomington	BDSC:34736;
Drosophila Stock	FlyBase:FBst0034736
Center	;
Bloomington Drosophila Stock Center	BDSC:34737; FlyBase:FBst0034737;
Bloomington	BDSC:34739;
Drosophila Stock	FlyBase:FBst0034739
Center	;
Bloomington	BDSC:34775;
Drosophila Stock	FlyBase:FBst0034775
Center	;
Bloomington Drosophila Stock Center	BDSC:34777; FlyBase:FBst0034777;
Bloomington	BDSC:34795;
Drosophila Stock	FlyBase:FBst0034795
Center	;
Bloomington Drosophila Stock Center	BDSC:34807; FlyBase:FBst0034807;
Bloomington	BDSC:34814;
Drosophila Stock	FlyBase:FBst0034814
Center	;
Bloomington	BDSC:34833;
Drosophila Stock	FlyBase:FBst0034833
Center	;
Bloomington	BDSC:34843;
Drosophila Stock	FlyBase:FBst0034843
Center	;
Bloomington	BDSC:34855;
Drosophila Stock	FlyBase:FBst0034855
Center	;
	Drosophila Stock Center Bloomington Drosophila Stock Center

Bloomington	BDSC:34862;
Drosophila Stock	FlyBase:FBst0034862
Center	;
Bloomington	BDSC:34869;
Drosophila Stock	FlyBase:FBst0034869
Center	;
Bloomington	BDSC:34881;
Drosophila Stock	FlyBase:FBst0034881
Center	;
Bloomington	BDSC:34884;
Drosophila Stock	FlyBase:FBst0034884
Center	;
Bloomington	BDSC:34894;
Drosophila Stock	FlyBase:FBst0034894
Center	;
Bloomington	BDSC:34898;
Drosophila Stock	FlyBase:FBst0034898
Center	;
Bloomington	BDSC:34900;
Drosophila Stock	FlyBase:FBst0034900
Center	;
Bloomington	BDSC:34910;
Drosophila Stock	FlyBase:FBst0034910
Center	;
Bloomington	BDSC:34929;
Drosophila Stock	FlyBase:FBst0034929
Center	;
Bloomington	BDSC:34938;
Drosophila Stock	FlyBase:FBst0034938
Center	;
Bloomington	BDSC:34958;
Drosophila Stock	FlyBase:FBst0034958
Center	;
	Drosophila Stock Center Bloomington Drosophila Stock Center

SC:34959; Base:FBst0034959
SC:34968; Base:FBst0034968
SC:34970; Base:FBst0034970
SC:34998; Base:FBst0034998
SC:35002; Base:FBst0035002
SC:35004; Base:FBst0035004
SC:35016; Base:FBst0035016
SC:35023; Base:FBst0035023
SC:35024; Base:FBst0035024
SC:35030; Base:FBst0035030
SC:35036; Base:FBst0035036
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D. melanogaster. Expresses dsRNA for RNAi of Cad99C (FBgn0039709) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01451}attP2	Bloomington Drosophila Stock Center	BDSC:35037; FlyBase:FBst0035037;
D. melanogaster. Expresses dsRNA for RNAi of pnt (FBgn0003118) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01452}attP2	Bloomington Drosophila Stock Center	BDSC:35038; FlyBase:FBst0035038
D. melanogaster. Expresses dsRNA for RNAi of numb (FBgn0002973) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01459}attP2	Bloomington Drosophila Stock Center	BDSC:35045; FlyBase:FBst0035045
D. melanogaster. Expresses dsRNA for RNAi of stan (FBgn0024836) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01464}attP2	Bloomington Drosophila Stock Center	BDSC:35050; FlyBase:FBst0035050 ;
D. melanogaster. Expresses dsRNA for RNAi of dco (FBgn0002413) under UAS control in the VALIUM22 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.GL00001}attP2/TM3, Sb[1]	Bloomington Drosophila Stock Center	BDSC:35134; FlyBase:FBst0035134 ;
D. melanogaster. Expresses dsRNA for RNAi of CkIlalpha (FBgn0264492) under UAS control in the VALIUM22 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.GL00003}attP2	Bloomington Drosophila Stock Center	BDSC:35136; FlyBase:FBst0035136 ;
D. melanogaster. Expresses dsRNA for RNAi of gish (FBgn0250823) under UAS control in the VALIUM22 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.GL00005}attP2	Bloomington Drosophila Stock Center	BDSC:35138; FlyBase:FBst0035138 ;
D. melanogaster. Expresses dsRNA for RNAi of Mkk4 (FBgn0024326) under UAS control in the VALIUM22 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.GL00010}attP2	Bloomington Drosophila Stock Center	BDSC:35143; FlyBase:FBst0035143 ;
D. melanogaster. Expresses dsRNA for RNAi of Cklalpha (FBgn0015024) under UAS control in the VALIUM22 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.GL00021}attP2	Bloomington Drosophila Stock Center	BDSC:35153; FlyBase:FBst0035153
D. melanogaster. Expresses dsRNA for RNAi of lic (FBgn0261524) under UAS control in the VALIUM22 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.GL00022}attP2/TM3, Sb[1]	Bloomington Drosophila Stock Center	BDSC:35154; FlyBase:FBst0035154 ;
D. melanogaster. Expresses dsRNA for RNAi of Btk29A (FBgn0003502) under UAS control in the VALIUM22 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.GL00027}attP2	Bloomington Drosophila Stock Center	BDSC:35159; FlyBase:FBst0035159

Bloomington	BDSC:35161;
Drosophila Stock	FlyBase:FBst0035161
Center	;
Bloomington	BDSC:35169;
Drosophila Stock	FlyBase:FBst0035169
Center	;
Bloomington	BDSC:35176;
Drosophila Stock	FlyBase:FBst0035176
Center	;
Bloomington	BDSC:35186;
Drosophila Stock	FlyBase:FBst0035186
Center	;
Bloomington Drosophila Stock Center	BDSC:35187; FlyBase:FBst0035187
Bloomington	BDSC:35195;
Drosophila Stock	FlyBase:FBst0035195
Center	;
Bloomington	BDSC:35210;
Drosophila Stock	FlyBase:FBst0035210
Center	;
Bloomington Drosophila Stock Center	BDSC:35211; FlyBase:FBst0035211;
Bloomington	BDSC:35225;
Drosophila Stock	FlyBase:FBst0035225
Center	;
Bloomington	BDSC:35238;
Drosophila Stock	FlyBase:FBst0035238
Center	;
Bloomington	BDSC:35244;
Drosophila Stock	FlyBase:FBst0035244
Center	;
	Drosophila Stock Center Bloomington Drosophila Stock Center

D. melanogaster. Expresses dsRNA for RNAi of p38b (FBgn0024846) under UAS control in the VALIUM22 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] Drosophila Stock V[+t1.8]=TRiP.GL00140} attP2 Center ; D. melanogaster. Expresses dsRNA for RNAi of app (FBgn0260941) under UAS control in the VALIUM22 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] Bloomington BDSC:35280; PryBase:FBst0 FlyBase:FBst0 F	
(FBgn0260941) under UAS control in the VALIUM22 Bloomington BDSC:35280;	0035280
v[+t1.8]=TRiP.GL00181}attP2 Center ;	
D. melanogaster. Expresses dsRNA for RNAi of sdt (FBgn0261873) under UAS control in the VALIUM22 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7]} v[+t1.8]=TRiP.GL00193}attP2 Bloomington Drosophila Stock Center ;)035291
D. melanogaster. Expresses dsRNA for RNAi of aux (FBgn0037218) under UAS control in the VALIUM22 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] Drosophila Stock v[+t1.8]=TRiP.GL00213}attP2 Bloomington BDSC:35310; FlyBase:FBst0 Center ;)035310
D. melanogaster. Expresses dsRNA for RNAi of CaMKII (FBgn0264607) under UAS control in the VALIUM22 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] Drosophila Stock v[+t1.8]=TRiP.GL00237}attP2/TM3, Sb[1] Bloomington Drosophila Stock Center ;)035330
D. melanogaster. Expresses dsRNA for RNAi of Ask1 (FBgn0014006) under UAS control in the VALIUM22 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7]} v[+t1.8]=TRiP.GL00238}attP2 Bloomington Drosophila Stock Center ;)035331
D. melanogaster. Expresses dsRNA for RNAi of SAK (FBgn0026371) under UAS control in the VALIUM22 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] Drosophila Stock v[+t1.8]=TRiP.GL00244}attP2/TM3, Sb[1] Bloomington BDSC:35335; FlyBase:FBst0 center ;)035335
D. melanogaster. Expresses dsRNA for RNAi of sgg (FBgn0003371) under UAS control in the VALIUM22 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.GL00277}attP2 Bloomington Drosophila Stock Center ;)035364
D. melanogaster. Expresses dsRNA for RNAi of hop (FBgn0004864) under UAS control in the VALIUM22 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] Drosophila Stock V[+t1.8]=TRiP.GL00305}attP2 Bloomington BDSC:35386; PyBase:FBst0 Center ;)035386
D. melanogaster. Expresses dsRNA for RNAi of Mekk1 (FBgn0024329) under UAS control in the VALIUM22 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] Drosophila Stock V[+t1.8]=TRiP.GL00322}attP2 Bloomington BDSC:35402; FlyBase:FBst0 Center ;)035402
D. melanogaster. Expresses dsRNA for RNAi of aop (FBgn0000097) under UAS control in the VALIUM22 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] Drosophila Stock V[+t1.8]=TRiP.GL00324}attP2 Bloomington BDSC:35404; PlyBase:FBst0 Center ;)035404

D. melanogaster. Expresses dsRNA for RNAi of Ras85D (FBgn0003205) under UAS control in the VALIUM22 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.GL00336}attP2/TM3, Sb[1]	Bloomington Drosophila Stock Center	BDSC:35414; FlyBase:FBst0035414;
D. melanogaster. Expresses dsRNA for RNAi of Nap1 (FBgn0015268) under UAS control in the VALIUM22 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.GL00370}attP2	Bloomington Drosophila Stock Center	BDSC:35445; FlyBase:FBst0035445 ;
D. melanogaster. Expresses dsRNA for RNAi of osa (FBgn0261885) under UAS control in the VALIUM22 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.GL00372}attP2	Bloomington Drosophila Stock Center	BDSC:35447; FlyBase:FBst0035447 ;
D. melanogaster. Expresses dsRNA for RNAi of key (FBgn0041205) under UAS control in the VALIUM22 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.GL00088}attP2	Bloomington Drosophila Stock Center	BDSC:35572; FlyBase:FBst0035572 ;
D. melanogaster. Expresses dsRNA for RNAi of Stat92E (FBgn0016917) under UAS control in the VALIUM22 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.GL00437}attP40/CyO	Bloomington Drosophila Stock Center	BDSC:35600; FlyBase:FBst0035600 ;
D. melanogaster. Expresses dsRNA for RNAi of Gcn5 (FBgn0020388) under UAS control in the VALIUM22 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.GL00439}attP40	Bloomington Drosophila Stock Center	BDSC:35601; FlyBase:FBst0035601 ;
D. melanogaster. Expresses dsRNA for RNAi of TER94 (FBgn0261014) under UAS control in the VALIUM22 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.GL00448}attP2	Bloomington Drosophila Stock Center	BDSC:35608; FlyBase:FBst0035608 ;
D. melanogaster. Expresses dsRNA for RNAi of wek (FBgn0001990) under UAS control in the VALIUM21 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.GLV21045}attP2	Bloomington Drosophila Stock Center	BDSC:35680; FlyBase:FBst0035680 ;
D. melanogaster. Expresses dsRNA for RNAi of bab2 (FBgn0025525) under UAS control in the VALIUM21 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.GLV21085}attP2	Bloomington Drosophila Stock Center	BDSC:35720; FlyBase:FBst0035720 ;
D. melanogaster. Expresses dsRNA for RNAi of Patj (FBgn0067864) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01489}attP2	Bloomington Drosophila Stock Center	BDSC:35747; FlyBase:FBst0035747
D. melanogaster. Expresses dsRNA for RNAi of Mtl (FBgn0039532) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01500}attP2	Bloomington Drosophila Stock Center	BDSC:35754; FlyBase:FBst0035754 ;

D. melanogaster. Expresses dsRNA for RNAi of gro (FBgn0001139) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01506}attP2	Bloomington Drosophila Stock Center	BDSC:35759; FlyBase:FBst0035759
D. melanogaster. Expresses dsRNA for RNAi of Wnt6 (FBgn0031902) under UAS control in the VALIUM22 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.GL00457}attP2	Bloomington Drosophila Stock Center	BDSC:35808; FlyBase:FBst0035808 ;
D. melanogaster. Expresses dsRNA for RNAi of CycE (FBgn0010382) under UAS control in the VALIUM22 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.GL00511}attP40	Bloomington Drosophila Stock Center	BDSC:36092; FlyBase:FBst0036092 ;
D. melanogaster. Expresses dsRNA for RNAi of Myd88 (FBgn0033402) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00183}attP2	Bloomington Drosophila Stock Center	BDSC:36107; FlyBase:FBst0036107
D. melanogaster. Expresses dsRNA for RNAi of Cul1 (FBgn0015509) under UAS control in the VALIUM22 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.GL00561}attP2	Bloomington Drosophila Stock Center	BDSC:36601; FlyBase:FBst0036601
D. melanogaster. Expresses dsRNA for RNAi of chico (FBgn0024248) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01553}attP2/TM3, Sb[1]	Bloomington Drosophila Stock Center	BDSC:36665; FlyBase:FBst0036665;
D. melanogaster. Expresses dsRNA for RNAi of Rbf2 (FBgn0038390) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01586}attP2	Bloomington Drosophila Stock Center	BDSC:36697; FlyBase:FBst0036697
D. melanogaster. Expresses dsRNA for RNAi of rictor (FBgn0031006) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01588}attP2	Bloomington Drosophila Stock Center	BDSC:36699; FlyBase:FBst0036699
D. melanogaster. Expresses dsRNA for RNAi of sty (FBgn0014388) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01599}attP2	Bloomington Drosophila Stock Center	BDSC:36709; FlyBase:FBst0036709
D. melanogaster. Expresses dsRNA for RNAi of Zyx (FBgn0011642) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01606}attP40	Bloomington Drosophila Stock Center	BDSC:36716; FlyBase:FBst0036716 ;
D. melanogaster. Expresses dsRNA for RNAi of Wnt2 (FBgn0004360) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01613}attP2	Bloomington Drosophila Stock Center	BDSC:36722; FlyBase:FBst0036722 ;

D. melanogaster. Expresses dsRNA for RNAi of Rbf		
(FBgn0015799) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS03004}attP2/TM3, Sb[1]	Bloomington Drosophila Stock Center	BDSC:36744; FlyBase:FBst0036744 ;
D. melanogaster. Expresses dsRNA for RNAi of dlg1 (FBgn0001624) under UAS control in the VALIUM20 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.JF02287}attP2	Bloomington Drosophila Stock Center	BDSC:36771; FlyBase:FBst0036771
D. melanogaster. Expresses dsRNA for RNAi of dpp (FBgn0000490) under UAS control in the VALIUM20 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.JF02455}attP2	Bloomington Drosophila Stock Center	BDSC:36779; FlyBase:FBst0036779
D. melanogaster. Expresses dsRNA for RNAi of DI (FBgn0000463) under UAS control in the VALIUM22 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.GL00520}attP40	Bloomington Drosophila Stock Center	BDSC:36784; FlyBase:FBst0036784 ;
D. melanogaster. Expresses dsRNA for RNAi of HDAC1 (FBgn0015805) under UAS control in the VALIUM22 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.GL01005}attP40	Bloomington Drosophila Stock Center	BDSC:36800; FlyBase:FBst0036800 ;
D. melanogaster. Expresses dsRNA for RNAi of Su(dx) (FBgn0003557) under UAS control in the VALIUM22 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.GL01077}attP2	Bloomington Drosophila Stock Center	BDSC:36836; FlyBase:FBst0036836 ;
D. melanogaster. Expresses dsRNA for RNAi of pbl (FBgn0003041) under UAS control in the VALIUM22 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.GL01092}attP2	Bloomington Drosophila Stock Center	BDSC:36841; FlyBase:FBst0036841 ;
D. melanogaster. Expresses dsRNA for RNAi of Sod2 (FBgn0010213) under UAS control in the VALIUM22 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.GL01015}attP40	Bloomington Drosophila Stock Center	BDSC:36871; FlyBase:FBst0036871
D. melanogaster. Expresses dsRNA for RNAi of Art1 (FBgn0037834) under UAS control in the VALIUM22 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.GL01072}attP2/TM3, Sb[1]	Bloomington Drosophila Stock Center	BDSC:36891; FlyBase:FBst0036891 ;
D. melanogaster. Expresses dsRNA for RNAi of CG10924 (FBgn0034356) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00200}attP2	Bloomington Drosophila Stock Center	BDSC:36915; FlyBase:FBst0036915
D. melanogaster. Expresses dsRNA for RNAi of trr (FBgn0023518) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01019}attP2	Bloomington Drosophila Stock Center	BDSC:36916; FlyBase:FBst0036916 ;

Bloomington Drosophila Stock Center	BDSC:37477; FlyBase:FBst0037477
Bloomington	BDSC:37489;
Drosophila Stock	FlyBase:FBst0037489
Center	;
Bloomington	BDSC:37520;
Drosophila Stock	FlyBase:FBst0037520
Center	;
Bloomington	BDSC:38194;
Drosophila Stock	FlyBase:FBst0038194
Center	;
Bloomington	BDSC:38208;
Drosophila Stock	FlyBase:FBst0038208
Center	;
Bloomington	BDSC:38209;
Drosophila Stock	FlyBase:FBst0038209
Center	;
Bloomington	BDSC:38215;
Drosophila Stock	FlyBase:FBst0038215
Center	;
Bloomington	BDSC:38249;
Drosophila Stock	FlyBase:FBst0038249
Center	;
Bloomington	BDSC:38276;
Drosophila Stock	FlyBase:FBst0038276
Center	;
Bloomington	BDSC:38286;
Drosophila Stock	FlyBase:FBst0038286
Center	;
Bloomington	BDSC:38373;
Drosophila Stock	FlyBase:FBst0038373
Center	;
	Drosophila Stock Center Bloomington Drosophila Stock Center

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D. melanogaster. Expresses dsRNA for RNAi of tws (FBgn0004889) under UAS control in the VALIUM22 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.GL00670}attP40/CyO	Bloomington Drosophila Stock Center	BDSC:38899; FlyBase:FBst0038899
D. melanogaster. Expresses dsRNA for RNAi of S (FBgn0003310) under UAS control in the VALIUM22 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.GL00686}attP2	Bloomington Drosophila Stock Center	BDSC:38914; FlyBase:FBst0038914 ;
D. melanogaster. Expresses dsRNA for RNAi of Pvf3 (FBgn0085407) under UAS control in the VALIUM20 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01876}attP40/CyO	Bloomington Drosophila Stock Center	BDSC:38962; FlyBase:FBst0038962 ;
D. melanogaster. Expresses dsRNA for RNAi of CanB2 (FBgn0015614) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01886}attP2	Bloomington Drosophila Stock Center	BDSC:38971; FlyBase:FBst0038971
D. melanogaster. Expresses dsRNA for RNAi of Pi3K21B (FBgn0020622) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01907}attP40	Bloomington Drosophila Stock Center	BDSC:38991; FlyBase:FBst0038991 ;
D. melanogaster. Expresses dsRNA for RNAi of par-6 (FBgn0026192) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01928}attP40	Bloomington Drosophila Stock Center	BDSC:39010; FlyBase:FBst0039010
D. melanogaster. Expresses dsRNA for RNAi of Pvf1 (FBgn0030964) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01958}attP40	Bloomington Drosophila Stock Center	BDSC:39038; FlyBase:FBst0039038
D. melanogaster. Expresses dsRNA for RNAi of Pka-C3 (FBgn0000489) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01970}attP2	Bloomington Drosophila Stock Center	BDSC:39050; FlyBase:FBst0039050
D. melanogaster. Expresses dsRNA for RNAi of DAAM (FBgn0025641) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01978}attP2	Bloomington Drosophila Stock Center	BDSC:39058; FlyBase:FBst0039058
D. melanogaster. Expresses dsRNA for RNAi of scrib (FBgn0263289) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01993}attP40/CyO	Bloomington Drosophila Stock Center	BDSC:39073; FlyBase:FBst0039073
D. melanogaster. Expresses dsRNA for RNAi of sina (FBgn0003410) under UAS control in the VALIUM20 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.HMS02008}attP40	Bloomington Drosophila Stock Center	BDSC:40842; FlyBase:FBst0040842 ;

Bloomington Drosophila Stock Center	BDSC:40871; FlyBase:FBst0040871
Bloomington Drosophila Stock Center	BDSC:40872; FlyBase:FBst0040872 ;
Bloomington Drosophila Stock Center	BDSC:40905; FlyBase:FBst0040905 ;
Bloomington Drosophila Stock Center	BDSC:40915; FlyBase:FBst0040915;
Bloomington Drosophila Stock Center	BDSC:41588; FlyBase:FBst0041588 ;
Bloomington Drosophila Stock Center	BDSC:41593; FlyBase:FBst0041593
Bloomington Drosophila Stock Center	BDSC:41598; FlyBase:FBst0041598 ;
Bloomington Drosophila Stock Center	BDSC:41605; FlyBase:FBst0041605
Bloomington Drosophila Stock Center	BDSC:41638; FlyBase:FBst0041638 ;
Bloomington Drosophila Stock Center	BDSC:41670; FlyBase:FBst0041670
Bloomington Drosophila Stock Center	BDSC:41699; FlyBase:FBst0041699
	Drosophila Stock Center Bloomington Drosophila Stock Center

Bloomington	BDSC:41702;
Drosophila Stock	FlyBase:FBst0041702
Center	;
Bloomington	BDSC:41723;
Drosophila Stock	FlyBase:FBst0041723
Center	;
Bloomington	BDSC:41823;
Drosophila Stock	FlyBase:FBst0041823
Center	;
Bloomington	BDSC:41830;
Drosophila Stock	FlyBase:FBst0041830
Center	;
Bloomington	BDSC:41878;
Drosophila Stock	FlyBase:FBst0041878
Center	;
Bloomington	BDSC:41904;
Drosophila Stock	FlyBase:FBst0041904
Center	;
Bloomington Drosophila Stock Center	BDSC:41908; FlyBase:FBst0041908
Bloomington	BDSC:41935;
Drosophila Stock	FlyBase:FBst0041935
Center	;
Bloomington	BDSC:41960;
Drosophila Stock	FlyBase:FBst0041960
Center	;
Bloomington	BDSC:41961;
Drosophila Stock	FlyBase:FBst0041961
Center	;
Bloomington Drosophila Stock Center	BDSC:41979; FlyBase:FBst0041979
	Drosophila Stock Center Bloomington Drosophila Stock Center

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D. melanogaster. Expresses dsRNA for RNAi of lgs (FBgn0039907) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS02381}attP2	Bloomington Drosophila Stock Center	BDSC:41983; FlyBase:FBst0041983
D. melanogaster. Expresses dsRNA for RNAi of Actbeta (FBgn0024913) under UAS control in the VALIUM20 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.HMJ02057}attP40	Bloomington Drosophila Stock Center	BDSC:42493; FlyBase:FBst0042493
D. melanogaster. Expresses dsRNA for RNAi of CG5059 (FBgn0037007) under UAS control in the VALIUM20 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.HMJ02058}attP40	Bloomington Drosophila Stock Center	BDSC:42494; FlyBase:FBst0042494
D. melanogaster. Expresses dsRNA for RNAi of msn (FBgn0010909) under UAS control in the VALIUM20 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.HMJ02084}attP40	Bloomington Drosophila Stock Center	BDSC:42518; FlyBase:FBst0042518 ;
D. melanogaster. Expresses dsRNA for RNAi of Rassf (FBgn0039055) under UAS control in the VALIUM20 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.HMJ02102}attP40	Bloomington Drosophila Stock Center	BDSC:42534; FlyBase:FBst0042534 ;
D. melanogaster. Expresses dsRNA for RNAi of sax (FBgn0003317) under UAS control in the VALIUM20 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.HMJ02118}attP40/CyO	Bloomington Drosophila Stock Center	BDSC:42546; FlyBase:FBst0042546 ;
D. melanogaster. Expresses dsRNA for RNAi of et (FBgn0031055) under UAS control in the VALIUM20 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.HMJ02213}attP40	Bloomington Drosophila Stock Center	BDSC:42557; FlyBase:FBst0042557
D. melanogaster. Expresses dsRNA for RNAi of nmo (FBgn0011817) under UAS control in the VALIUM20 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.HMJ02229}attP40	Bloomington Drosophila Stock Center	BDSC:42570; FlyBase:FBst0042570
D. melanogaster. Expresses dsRNA for RNAi of CG12147 (FBgn0037325) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS02447}attP2	Bloomington Drosophila Stock Center	BDSC:42612; FlyBase:FBst0042612 ;
D. melanogaster. Expresses dsRNA for RNAi of Act5C (FBgn0000042) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS02487}attP2	Bloomington Drosophila Stock Center	BDSC:42651; FlyBase:FBst0042651
D. melanogaster. Expresses dsRNA for RNAi of Act87E (FBgn0000046) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS02488}attP2/TM3, Sb[1]	Bloomington Drosophila Stock Center	BDSC:42652; FlyBase:FBst0042652 ;

D. melanogaster. Expresses dsRNA for RNAi of emc (FBgn0000575) under UAS control in the VALIUM22 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.GL00724}attP2	Bloomington Drosophila Stock Center	BDSC:42768; FlyBase:FBst0042768
D. melanogaster. Expresses dsRNA for RNAi of the Stellate gene family (FBgn0003523) plus Ste12DOR and SteXh:CG42398 (FBgn0044817 and FBgn0259817) under UAS control in the VALIUM22 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.GL01156}attP2	Bloomington Drosophila Stock Center	BDSC:42786; FlyBase:FBst0042786 ;
D. melanogaster. Expresses dsRNA for RNAi of Socs44A (FBgn0033266) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS02515}attP2	Bloomington Drosophila Stock Center	BDSC:42830; FlyBase:FBst0042830 ;
D. melanogaster. Expresses dsRNA for RNAi of gcm (FBgn0014179) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS02582}attP40	Bloomington Drosophila Stock Center	BDSC:42889; FlyBase:FBst0042889
D. melanogaster. Expresses dsRNA for RNAi of CkIlbeta (FBgn0000259) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS02636}attP40	Bloomington Drosophila Stock Center	BDSC:42943; FlyBase:FBst0042943
D. melanogaster. Expresses dsRNA for RNAi of smo (FBgn0003444) under UAS control in the VALIUM22 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.GL01472}attP2	Bloomington Drosophila Stock Center	BDSC:43134; FlyBase:FBst0043134 ;
D. melanogaster. Expresses dsRNA for RNAi of dock (FBgn0010583) under UAS control in the VALIUM22 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.GL01519}attP2/TM3, Sb[1]	Bloomington Drosophila Stock Center	BDSC:43176; FlyBase:FBst0043176 ;
D. melanogaster. Expresses dsRNA for RNAi of Mad (FBgn0011648) under UAS control in the VALIUM22 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.GL01527}attP40	Bloomington Drosophila Stock Center	BDSC:43183; FlyBase:FBst0043183
D. melanogaster. Expresses dsRNA for RNAi of Pp2A-29B (FBgn0260439) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01921}attP2/TM3, Sb[1]	Bloomington Drosophila Stock Center	BDSC:43283; FlyBase:FBst0043283 ;
D. melanogaster. Expresses dsRNA for RNAi of CG11658 (FBgn0036196) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS02671}attP40	Bloomington Drosophila Stock Center	BDSC:43298; FlyBase:FBst0043298 ;

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D. melanogaster. Expresses dsRNA for RNAi of Myc (FBgn0262656) under UAS control in the VALIUM22 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.GL01314}attP40	Bloomington Drosophila Stock Center	BDSC:43962; FlyBase:FBst0043962 ;
D. melanogaster. Expresses dsRNA for RNAi of pav (FBgn0011692) under UAS control in the VALIUM22 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.GL01316}attP40	Bloomington Drosophila Stock Center	BDSC:43963; FlyBase:FBst0043963
D. melanogaster. Expresses dsRNA for RNAi of eIF4EHP (FBgn0053100) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS02703}attP40	Bloomington Drosophila Stock Center	BDSC:43990; FlyBase:FBst0043990 ;
D. melanogaster. Expresses dsRNA for RNAi of Atg1 (FBgn0260945) under UAS control in the VALIUM20 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.HMS02750}attP40	Bloomington Drosophila Stock Center	BDSC:44034; FlyBase:FBst0044034
D. melanogaster. Expresses dsRNA for RNAi of tefu (FBgn0045035) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS02790}attP40	Bloomington Drosophila Stock Center	BDSC:44073; FlyBase:FBst0044073
D. melanogaster. Expresses dsRNA for RNAi of bi (FBgn0000179) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS02815}attP2	Bloomington Drosophila Stock Center	BDSC:44095; FlyBase:FBst0044095
D. melanogaster. Expresses dsRNA for RNAi of mad2 (FBgn0035640) under UAS control in the VALIUM22 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.GLC01381}attP2	Bloomington Drosophila Stock Center	BDSC:44430; FlyBase:FBst0044430 ;
D. melanogaster. Expresses dsRNA for RNAi of ebi (FBgn0263933) under UAS control in the VALIUM22 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.GLC01413}attP40	Bloomington Drosophila Stock Center	BDSC:44443; FlyBase:FBst0044443;
D. melanogaster. Expresses dsRNA for RNAi of dx (FBgn0000524) under UAS control in the VALIUM22 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.GLC01607}attP2	Bloomington Drosophila Stock Center	BDSC:44455; FlyBase:FBst0044455;
D. melanogaster. Expresses dsRNA for RNAi of fz3 (FBgn0027343) under UAS control in the VALIUM22 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.GLC01626}attP2	Bloomington Drosophila Stock Center	BDSC:44468; FlyBase:FBst0044468
D. melanogaster. Expresses dsRNA for RNAi of cos (FBgn0000352) under UAS control in the VALIUM20 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.HMC02347}attP2	Bloomington Drosophila Stock Center	BDSC:44472; FlyBase:FBst0044472

Bloomington Drosophila Stock Center	BDSC:44633; FlyBase:FBst0044633
Bloomington Drosophila Stock Center	BDSC:50529; FlyBase:FBst0050529
Bloomington Drosophila Stock Center	BDSC:50540; FlyBase:FBst0050540
Bloomington Drosophila Stock Center	BDSC:50594; FlyBase:FBst0050594 ;
Bloomington Drosophila Stock Center	BDSC:50610; FlyBase:FBst0050610
Bloomington Drosophila Stock Center	BDSC:50625; FlyBase:FBst0050625
Bloomington Drosophila Stock Center	BDSC:50663; FlyBase:FBst0050663
Bloomington Drosophila Stock Center	BDSC:50712; FlyBase:FBst0050712 ;
Bloomington Drosophila Stock Center	BDSC:50730; FlyBase:FBst0050730 ;
Bloomington Drosophila Stock Center	BDSC:50951; FlyBase:FBst0050951
Bloomington Drosophila Stock Center	BDSC:50972; FlyBase:FBst0050972
	Drosophila Stock Center Bloomington Drosophila Stock Center

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D. melanogaster. Expresses dsRNA for RNAi of dpn (FBgn0010109) under UAS control in the VALIUM20 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.HMC03154}attP2	Bloomington Drosophila Stock Center	BDSC:51440; FlyBase:FBst0051440
D. melanogaster. Expresses dsRNA for RNAi of Spn27A (FBgn0028990) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMC03159}attP2	Bloomington Drosophila Stock Center	BDSC:51445; FlyBase:FBst0051445 ;
D. melanogaster. Expresses dsRNA for RNAi of ttv (FBgn0265974) under UAS control in the VALIUM20 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.HMC03225}attP40	Bloomington Drosophila Stock Center	BDSC:51480; FlyBase:FBst0051480
D. melanogaster. Expresses dsRNA for RNAi of Mipp2 (FBgn0026060) under UAS control in the VALIUM20 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.HMC03229}attP40/CyO	Bloomington Drosophila Stock Center	BDSC:51482; FlyBase:FBst0051482 ;
D. melanogaster. Expresses dsRNA for RNAi of kibra (FBgn0262127) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMC03256}attP2	Bloomington Drosophila Stock Center	BDSC:51499; FlyBase:FBst0051499
D. melanogaster. Expresses dsRNA for RNAi of tld (FBgn0003719) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMC03275}attP2	Bloomington Drosophila Stock Center	BDSC:51507; FlyBase:FBst0051507
D. melanogaster. Expresses dsRNA for RNAi of S6kII (FBgn0262866) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMC03140}attP40	Bloomington Drosophila Stock Center	BDSC:51694; FlyBase:FBst0051694 ;
D. melanogaster. Expresses dsRNA for RNAi of ast (FBgn0015905) under UAS control in the VALIUM20 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.HMC03173}attP2	Bloomington Drosophila Stock Center	BDSC:51700; FlyBase:FBst0051700
D. melanogaster. Expresses dsRNA for RNAi of ras (FBgn0003204) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMC03250}attP2	Bloomington Drosophila Stock Center	BDSC:51717; FlyBase:FBst0051717
D. melanogaster. Expresses dsRNA for RNAi of E(spl)mgamma-HLH (FBgn0002735) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMC03315}attP2	Bloomington Drosophila Stock Center	BDSC:51762; FlyBase:FBst0051762 ;
D. melanogaster. Expresses dsRNA for RNAi of Src64B (FBgn0262733) under UAS control in the VALIUM20 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.HMC03327}attP40	Bloomington Drosophila Stock Center	BDSC:51772; FlyBase:FBst0051772 ;

Bloomington Drosophila Stock Center	BDSC:51789; FlyBase:FBst0051789
Bloomington	BDSC:52883;
Drosophila Stock	FlyBase:FBst0052883
Center	;
Bloomington	BDSC:52908;
Drosophila Stock	FlyBase:FBst0052908
Center	;
Bloomington	BDSC:52931;
Drosophila Stock	FlyBase:FBst0052931
Center	;
Bloomington Drosophila Stock Center	BDSC:53003; FlyBase:FBst0053003
Bloomington	BDSC:53342;
Drosophila Stock	FlyBase:FBst0053342
Center	;
Bloomington	BDSC:53349;
Drosophila Stock	FlyBase:FBst0053349
Center	;
Bloomington Drosophila Stock Center	BDSC:53697; FlyBase:FBst0053697
Bloomington	BDSC:53704;
Drosophila Stock	FlyBase:FBst0053704
Center	;
Bloomington	BDSC:53880;
Drosophila Stock	FlyBase:FBst0053880
Center	;
Bloomington	BDSC:53884;
Drosophila Stock	FlyBase:FBst0053884
Center	;
	Drosophila Stock Center Bloomington Drosophila Stock Center

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Bloomington	BDSC:53890;
Drosophila Stock	FlyBase:FBst0053890
Center	;
Bloomington	BDSC:54851;
Drosophila Stock	FlyBase:FBst0054851
Center	;
Bloomington Drosophila Stock Center	BDSC:55169; FlyBase:FBst0055169
Bloomington	BDSC:55260;
Drosophila Stock	FlyBase:FBst0055260
Center	;
Bloomington	BDSC:55276;
Drosophila Stock	FlyBase:FBst0055276
Center	;
Bloomington	BDSC:55302;
Drosophila Stock	FlyBase:FBst0055302
Center	;
Bloomington	BDSC:55318;
Drosophila Stock	FlyBase:FBst0055318
Center	;
Bloomington	BDSC:55352;
Drosophila Stock	FlyBase:FBst0055352
Center	;
Bloomington	BDSC:55367;
Drosophila Stock	FlyBase:FBst0055367
Center	;
Bloomington Drosophila Stock Center	BDSC:55379; FlyBase:FBst0055379
Bloomington	BDSC:55404;
Drosophila Stock	FlyBase:FBst0055404
Center	;
	Drosophila Stock Center Bloomington Drosophila Stock Center

Bloomington Drosophila Stock Center	BDSC:55679; FlyBase:FBst0055679
Bloomington Drosophila Stock Center	BDSC:55681; FlyBase:FBst0055681 ;
Bloomington Drosophila Stock Center	BDSC:55686; FlyBase:FBst0055686 ;
Bloomington Drosophila Stock Center	BDSC:55859; FlyBase:FBst0055859 ;
Bloomington Drosophila Stock Center	BDSC:55866; FlyBase:FBst0055866 ;
Bloomington Drosophila Stock Center	BDSC:55867; FlyBase:FBst0055867
Bloomington Drosophila Stock Center	BDSC:55871; FlyBase:FBst0055871
Bloomington Drosophila Stock Center	BDSC:55874; FlyBase:FBst0055874 ;
Bloomington Drosophila Stock Center	BDSC:55893; FlyBase:FBst0055893
Bloomington Drosophila Stock Center	BDSC:55899; FlyBase:FBst0055899
Bloomington Drosophila Stock Center	BDSC:55903; FlyBase:FBst0055903 ;
	Drosophila Stock Center Bloomington Drosophila Stock Center

Bloomington Drosophila Stock Center	BDSC:55908; FlyBase:FBst0055908 ;
Bloomington Drosophila Stock Center	BDSC:55921; FlyBase:FBst0055921 ;
Bloomington Drosophila Stock Center	BDSC:55926; FlyBase:FBst0055926 ;
Bloomington Drosophila Stock Center	BDSC:57307; FlyBase:FBst0057307
Bloomington Drosophila Stock Center	BDSC:57497; FlyBase:FBst0057497
Bloomington Drosophila Stock Center	BDSC:57867; FlyBase:FBst0057867
Bloomington Drosophila Stock Center	BDSC:58067; FlyBase:FBst0058067 ;
Bloomington Drosophila Stock Center	BDSC:58309; FlyBase:FBst0058309 ;
Bloomington Drosophila Stock Center	BDSC:58499; FlyBase:FBst0058499
Bloomington Drosophila Stock Center	BDSC:62372; FlyBase:FBst0062372 ;
Bloomington Drosophila Stock Center	BDSC:62513; FlyBase:FBst0062513
	Drosophila Stock Center Bloomington Drosophila Stock Center

D. melanogaster. Expresses dsRNA for RNAi of CG2199 (FBgn0035213) under UAS control in the VALIUM20 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.HMJ30228}attP40	Bloomington Drosophila Stock Center	BDSC:63661; FlyBase:FBst0063661 ;
D. melanogaster. Expresses dsRNA for RNAi of tsr (FBgn0011726) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00534}attP2	Bloomington Drosophila Stock Center	BDSC:65055; FlyBase:FBst0065055;
D. melanogaster. Expresses dsRNA for RNAi of CG3630 (FBgn0023540) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMC06220}attP2	Bloomington Drosophila Stock Center	BDSC:65945; FlyBase:FBst0065945
D. melanogaster. Expresses dsRNA for RNAi of tub (FBgn0003882) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS05426}attP40	Bloomington Drosophila Stock Center	BDSC:66960; FlyBase:FBst0066960
D. melanogaster. Expresses dsRNA for RNAi of elF4E-6 (FBgn0039622) under UAS control.	Vienna Drosophila Resource Center	VDRC:v17580
D. melanogaster. Expresses dsRNA for RNAi of gd (FBgn0000808) under UAS control.	Vienna Drosophila Resource Center	VDRC:v14892
D. melanogaster. Expresses dsRNA for RNAi of Act88F (FBgn0000047) under UAS control.	Vienna Drosophila Resource Center	VDRC:v9780
D. melanogaster. Expresses dsRNA for RNAi of eRF1 (FBgn0036974) under UAS control.	Vienna Drosophila Resource Center	VDRC:v45027
D. melanogaster. Expresses dsRNA for RNAi of E(spl)m2-BFM (FBgn0002592) under UAS control.	Vienna Drosophila Resource Center	VDRC:v30115
D. melanogaster. Expresses dsRNA for RNAi of tsl (FBgn0003867) under UAS control.	Vienna Drosophila Resource Center	VDRC:v14430
D. melanogaster. Expresses dsRNA for RNAi of wbl (FBgn0004003) under UAS control.	Vienna Drosophila Resource Center	VDRC:v13864
D. melanogaster. Expresses dsRNA for RNAi of boss (FBgn0000206) under UAS control.	Vienna Drosophila Resource Center	VDRC:v4365
D. melanogaster. Expresses dsRNA for RNAi of CG32396 (FBgn0020251) under UAS control.	Vienna Drosophila Resource Center	VDRC:v41896
D. melanogaster. Expresses dsRNA for RNAi of Rac2 (FBgn0014011) under UAS control.	Vienna Drosophila Resource Center	VDRC:v28926
D. melanogaster. Expresses dsRNA for RNAi of ihog (FBgn0031872) under UAS control.	Vienna Drosophila Resource Center	VDRC:v29898
D. melanogaster. Expresses dsRNA for RNAi of sog (FBgn0003463) under UAS control.	Vienna Drosophila Resource Center	VDRC:v37405
D. melanogaster. Expresses dsRNA for RNAi of CG9314 (FBgn0032061) under UAS control.	Vienna Drosophila Resource Center	VDRC:v44647
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D. melanogaster. Expresses dsRNA for RNAi of sgl (FBgn0261445) under UAS control.	Vienna Drosophila Resource Center	VDRC:v29434
D. melanogaster. Expresses dsRNA for RNAi of mirr (FBgn0014343) under UAS control.	Vienna Drosophila Resource Center	VDRC:v50134
D. melanogaster. Expresses dsRNA for RNAi of eIF-4B (FBgn0020660) under UAS control.	Vienna Drosophila Resource Center	VDRC:v31364
D. melanogaster. Expresses dsRNA for RNAi of rasp (FBgn0024194) under UAS control.	Vienna Drosophila Resource Center	VDRC:v6459
D. melanogaster. Expresses dsRNA for RNAi of PGRP-SA (FBgn0030310) under UAS control.	Vienna Drosophila Resource Center	VDRC:v5594
D. melanogaster. Expresses dsRNA for RNAi of sinah (FBgn0259794) under UAS control.	Vienna Drosophila Resource Center	VDRC:v17118
D. melanogaster. Expresses dsRNA for RNAi of lft (FBgn0032230) under UAS control.	Vienna Drosophila Resource Center	VDRC:v32146
D. melanogaster. Expresses dsRNA for RNAi of pyr (FBgn0033649) under UAS control.	Vienna Drosophila Resource Center	VDRC:v36524
D. melanogaster. Expresses dsRNA for RNAi of Ssl (FBgn0015300) under UAS control.	Vienna Drosophila Resource Center	VDRC:v17282
D. melanogaster. Expresses dsRNA for RNAi of Pvf2 (FBgn0031888) under UAS control.	Vienna Drosophila Resource Center	VDRC:v7628
D. melanogaster. Expresses dsRNA for RNAi of spz4 (FBgn0032362) under UAS control.	Vienna Drosophila Resource Center	VDRC:v7679
D. melanogaster. Expresses dsRNA for RNAi of IM23 (FBgn0034328) under UAS control.	Vienna Drosophila Resource Center	VDRC:v15384
D. melanogaster. Expresses dsRNA for RNAi of botv (FBgn0027535) under UAS control.	Vienna Drosophila Resource Center	VDRC:v37186
D. melanogaster. Expresses dsRNA for RNAi of G6P (FBgn0031463) under UAS control.	Vienna Drosophila Resource Center	VDRC:v7261
D. melanogaster. Expresses dsRNA for RNAi of stet (FBgn0020248) under UAS control.	Vienna Drosophila Resource Center	VDRC:v7434
D. melanogaster. Expresses dsRNA for RNAi of RanBPM (FBgn0262114) under UAS control.	Vienna Drosophila Resource Center	VDRC:v45981
D. melanogaster. Expresses dsRNA for RNAi of Src42A (FBgn0264959) under UAS control.	Vienna Drosophila Resource Center	VDRC:v26019
D. melanogaster. Expresses dsRNA for RNAi of nkd (FBgn0002945) under UAS control.	Vienna Drosophila Resource Center	VDRC:v3004
D. melanogaster. Expresses dsRNA for RNAi of boi (FBgn0040388) under UAS control.	Vienna Drosophila Resource Center	VDRC:v3060
D. melanogaster. Expresses dsRNA for RNAi of CR45683; Tehao (FBgn0026760) under UAS control.	Vienna Drosophila Resource Center	VDRC:v17903
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D. melanogaster. Expresses dsRNA for RNAi of CG45087; Pepck (FBgn0003067) under UAS control.	Vienna Drosophila Resource Center	VDRC:v20529
D. melanogaster. Expresses dsRNA for RNAi of RpL13A (FBgn0037351) under UAS control.	Vienna Drosophila Resource Center	VDRC:v101369
D. melanogaster. Expresses dsRNA for RNAi of Usp7 (FBgn0030366) under UAS control.	Vienna Drosophila Resource Center	VDRC:v110324
D. melanogaster. Expresses dsRNA for RNAi of tsg (FBgn0003865) under UAS control.	Vienna Drosophila Resource Center	VDRC:v108750
D. melanogaster. Expresses dsRNA for RNAi of nec (FBgn0002930) under UAS control.	Vienna Drosophila Resource Center	VDRC:v108366
D. melanogaster. Expresses dsRNA for RNAi of Nle (FBgn0021874) under UAS control.	Vienna Drosophila Resource Center	VDRC:v110728
D. melanogaster. Expresses dsRNA for RNAi of Brd (FBgn0000216) under UAS control.	Vienna Drosophila Resource Center	VDRC:v107929
D. melanogaster. Expresses dsRNA for RNAi of Shc (FBgn0015296) under UAS control.	Vienna Drosophila Resource Center	VDRC:v103906
D. melanogaster. Expresses dsRNA for RNAi of Hs6st (FBgn0038755) under UAS control.	Vienna Drosophila Resource Center	VDRC:v110424
D. melanogaster. Expresses dsRNA for RNAi of fz4 (FBgn0027342) under UAS control.	Vienna Drosophila Resource Center	VDRC:v102339
D. melanogaster. Expresses dsRNA for RNAi of bib (FBgn0000180) under UAS control.	Vienna Drosophila Resource Center	VDRC:v103327
D. melanogaster. Expresses dsRNA for RNAi of Wnt10 (FBgn0031903) under UAS control.	Vienna Drosophila Resource Center	VDRC:v100867
D. melanogaster. Expresses dsRNA for RNAi of Tom (FBgn0026320) under UAS control.	Vienna Drosophila Resource Center	VDRC:v101652
D. melanogaster. Expresses dsRNA for RNAi of Pli (FBgn0025574) under UAS control.	Vienna Drosophila Resource Center	VDRC:v106776
D. melanogaster. Expresses dsRNA for RNAi of drk (FBgn0004638) under UAS control.	Vienna Drosophila Resource Center	VDRC:v105498
D. melanogaster. Expresses dsRNA for RNAi of por (FBgn0004957) under UAS control.	Vienna Drosophila Resource Center	VDRC:v100780
D. melanogaster. Expresses dsRNA for RNAi of wls (FBgn0036141) under UAS control.	Vienna Drosophila Resource Center	VDRC:v103812
D. melanogaster. Expresses dsRNA for RNAi of CG6843 (FBgn0036827) under UAS control.	Vienna Drosophila Resource Center	VDRC:v109411
D. melanogaster. Expresses dsRNA for RNAi of spz3 (FBgn0031959) under UAS control.	Vienna Drosophila Resource Center	VDRC:v102871
D. melanogaster. Expresses dsRNA for RNAi of kuz (FBgn0259984) under UAS control.	Vienna Drosophila Resource Center	VDRC:v107036
		

D. melanogaster. Expresses dsRNA for RNAi of Hs3st-B (FBgn0031005) under UAS control.	Vienna Drosophila Resource Center	VDRC:v110601
D. melanogaster. Expresses dsRNA for RNAi of Tace (FBgn0039734) under UAS control.	Vienna Drosophila Resource Center	VDRC:v106335
D. melanogaster. Expresses dsRNA for RNAi of eIF-1A (FBgn0026250) under UAS control.	Vienna Drosophila Resource Center	VDRC:v100611
D. melanogaster. Expresses dsRNA for RNAi of Socs16D (FBgn0030869) under UAS control.	Vienna Drosophila Resource Center	VDRC:v100568
D. melanogaster. Expresses dsRNA for RNAi of E(spl)malpha-BFM (FBgn0002732) under UAS control.	Vienna Drosophila Resource Center	VDRC:v109384
D. melanogaster. Expresses dsRNA for RNAi of E(spl)m6-BFM (FBgn0002632) under UAS control.	Vienna Drosophila Resource Center	VDRC:v101965
D. melanogaster. Expresses dsRNA for RNAi of VhaM8.9 (FBgn0037671) under UAS control.	Vienna Drosophila Resource Center	VDRC:v105281
D. melanogaster. Expresses dsRNA for RNAi of melt (FBgn0023001) under UAS control.	Vienna Drosophila Resource Center	VDRC:v105110
D. melanogaster. Expresses dsRNA for RNAi of SkpB (FBgn0026176) under UAS control.	Vienna Drosophila Resource Center	VDRC:v106521
D. melanogaster. Expresses dsRNA for RNAi of CkIlbeta2 (FBgn0026136) under UAS control.	Vienna Drosophila Resource Center	VDRC:v102633
D. melanogaster. Expresses dsRNA for RNAi of spz6 (FBgn0035056) under UAS control.	Vienna Drosophila Resource Center	VDRC:v100897
D. melanogaster. Expresses dsRNA for RNAi of Rok (FBgn0026181) under UAS control.	Vienna Drosophila Resource Center	VDRC:v104675
D. melanogaster. Expresses dsRNA for RNAi of CG9962 (FBgn0031441) under UAS control.	Vienna Drosophila Resource Center	VDRC:v108721
D. melanogaster. Expresses dsRNA for RNAi of spz5 (FBgn0035379) under UAS control.	Vienna Drosophila Resource Center	VDRC:v102389
D. melanogaster. Expresses dsRNA for RNAi of Act57B (FBgn0000044) under UAS control.	Vienna Drosophila Resource Center	VDRC:v102129
D. melanogaster. Expresses dsRNA for RNAi of ndl (FBgn0002926) under UAS control.	Vienna Drosophila Resource Center	VDRC:v102818
D. melanogaster. Expresses dsRNA for RNAi of vn (FBgn0003984) under UAS control.	Vienna Drosophila Resource Center	VDRC:v109437
D. melanogaster. Expresses dsRNA for RNAi of ECSIT (FBgn0028436) under UAS control.	Vienna Drosophila Resource Center	VDRC:v106141
D. melanogaster. Expresses dsRNA for RNAi of SkpE (FBgn0031074) under UAS control.	Vienna Drosophila Resource Center	VDRC:v109539
D. melanogaster. Expresses dsRNA for RNAi of SkpF (FBgn0034863) under UAS control.	Vienna Drosophila Resource Center	VDRC:v106572
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D. melanogaster. Expresses dsRNA for RNAi of kek1 (FBgn0015399) under UAS control.	Vienna Drosophila Resource Center	VDRC:v101166
D. melanogaster. Expresses dsRNA for RNAi of ths (FBgn0033652) under UAS control.	Vienna Drosophila Resource Center	VDRC:v102441
D. melanogaster. Expresses dsRNA for RNAi of Ilp8 (FBgn0036690) under UAS control.	Vienna Drosophila Resource Center	VDRC:v102604
D. melanogaster. Expresses dsRNA for RNAi of CG15800 (FBgn0034904) under UAS control.	Vienna Drosophila Resource Center	VDRC:v110049
D. melanogaster. Expresses dsRNA for RNAi of IM3 (FBgn0040736) under UAS control.	Vienna Drosophila Resource Center	VDRC:v104908
D. melanogaster. Expresses dsRNA for RNAi of Roc1a (FBgn0025638) under UAS control.	Vienna Drosophila Resource Center	VDRC:v106315
D. melanogaster. Expresses dsRNA for RNAi of dod (FBgn0015379) under UAS control.	Vienna Drosophila Resource Center	VDRC:v110593
D. melanogaster. Expresses dsRNA for RNAi of hipk (FBgn0035142) under UAS control.	Vienna Drosophila Resource Center	VDRC:v108254
D. melanogaster. Expresses dsRNA for RNAi of ave (FBgn0050476) under UAS control.	Vienna Drosophila Resource Center	VDRC:v101471
D. melanogaster. Expresses dsRNA for RNAi of boca (FBgn0004132) under UAS control.	Vienna Drosophila Resource Center	VDRC:v108406
D. melanogaster. Expresses dsRNA for RNAi of gskt (FBgn0046332) under UAS control.	Vienna Drosophila Resource Center	VDRC:v107429
D. melanogaster. Expresses dsRNA for RNAi of stumps (FBgn0020299) under UAS control.	Vienna Drosophila Resource Center	VDRC:v105603
D. melanogaster. Expresses dsRNA for RNAi of CG31431 (FBgn0051431) under UAS control.	Vienna Drosophila Resource Center	VDRC:v104697
D. melanogaster. Expresses dsRNA for RNAi of scw (FBgn0005590) under UAS control.	Vienna Drosophila Resource Center	VDRC:v105303
D. melanogaster. Expresses dsRNA for RNAi of fry (FBgn0016081) under UAS control.	Vienna Drosophila Resource Center	VDRC:v103569
D. melanogaster. Expresses dsRNA for RNAi of Krn (FBgn0052179) under UAS control.	Vienna Drosophila Resource Center	VDRC:v104299
D. melanogaster. Expresses dsRNA for RNAi of pxb (FBgn0053207) under UAS control.	Vienna Drosophila Resource Center	VDRC:v102240
D. melanogaster. Expresses dsRNA for RNAi of cv-c (FBgn0285955) under UAS control.	Vienna Drosophila Resource Center	VDRC:v105435
D. melanogaster. Expresses dsRNA for RNAi of cic (FBgn0262582) under UAS control.	Vienna Drosophila Resource Center	VDRC:v103805
D. melanogaster. Expresses dsRNA for RNAi of dia (FBgn0011202) under UAS control.	Vienna Drosophila Resource Center	VDRC:v103914

D. melanogaster. Expresses dsRNA for RNAi of SkpD;SkpC (FBgn0026174) under UAS control.	Vienna Drosophila Resource Center	VDRC:v109181
D. melanogaster. Expresses dsRNA for RNAi of drk (FBgn0004638) under UAS control.	Vienna Drosophila Resource Center	VDRC:v105498
D. melanogaster. Expresses dsRNA for RNAi of botv (FBgn0027535) under UAS control.	Vienna Drosophila Resource Center	VDRC:v37186

1045 Reagents

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Name	Catalogue number	Usage
Hoechst 33321	H1399 (ThermoFisher)	1ug/ml final concentration

References

- Ables, E.T., Drummond-Barbosa, D., 2017. Steroid Hormones and the Physiological Regulation
- of Tissue-Resident Stem Cells: Lessons from the *Drosophila* Ovary. Current Stem Cell Reports
- 1051 3, 9-18.

1048

- Albert, I., Albert, R., 2004. Conserved network motifs allow protein-protein interaction prediction.
- 1053 Bioinformatics 20, 3346-3352.
- Albert, R., Barabási, A.-L., 2002. Statistical mechanics of complex networks. Reviews of Modern
- 1055 Physics 74, 47-97.
- Allbee, A.W., Rincon-Limas, D.E., Biteau, B., 2018. *Lmx1a* is required for the development of
- the ovarian stem cell niche in *Drosophila*. Development 145, dev163394.
- Azevedo, R.B.R., French, V., Partridge, L., 1996. Thermal evolution of egg size in *Drosophila*
- 1059 *melanogaster*. Evolution 50, 2338-2345.
- 1060 Barabási, A.L., Albert, R., 1999. Emergence of scaling in random networks. Science 286, 509-
- 1061 512.
- Barabási, A.L., Oltvai, Z.N., 2004. Network biology: understanding the cell's functional
- organization. Nat. Rev. Genet. 5, 101-113.
- Barrett, K., Leptin, M., Settleman, J., 1997. The Rho GTPase and a putative RhoGEF mediate a
- signaling pathway for the cell shape changes in *Drosophila* gastrulation. Cell 91, 905-915.
- 1066 Berger, S.I., Posner, J.M., Ma'ayan, A., 2007. Genes2Networks: connecting lists of gene
- symbols using mammalian protein interactions databases. BMC Bioinformatics 8, 372.
- Bergland, A.O., Genissel, A., Nuzhdin, S.V., Tatar, M., 2008. Quantitative trait loci affecting
- phenotypic plasticity and the allometric relationship of ovariole number and thorax length in
- 1070 Drosophila melanogaster. Genetics 180, 567-582.
- 1071 Bolívar, J., Pearson, J., López-Onieva, L., González-Reyes, A., 2006. Genetic dissection of a
- stem cell niche: the case of the *Drosophila* ovary. Dev. Dyn. 235, 2969-2979.
- Bromberg, K.D., Ma'ayan, A., Neves, S.R., Iyengar, R., 2008. Design logic of a cannabinoid
- receptor signaling network that triggers neurite outgrowth. Science 320, 903-909.
- 1075 Capy, P., Pla, E., David, J.R., 1993. Phenotypic and genetic variability of morphometrical traits
- in natural populations of *Drosophila melanogaster* and *D. simulans*. I. Geographic variations.
- 1077 Genet. Sel. Evol. 25, 517-536.
- 1078 Capy, P., Pla, E., David, J.R., 1994. Phenotypic and genetic variability of morphometrical traits
- in natural populations of *Drosophila melanogaster* and *D simulans*. II. Within-population
- 1080 variability. Genetics Selection Evolution 26, 15-28.
- 1081 Cayirlioglu, P., Ward, W.O., Silver Key, S.C., Duronio, R.J., 2003. Transcriptional repressor
- 1082 functions of *Drosophila* E2F1 and E2F2 cooperate to inhibit genomic DNA synthesis in ovarian
- 1083 follicle cells. Mol. Cell. Biol. 23, 2123-2134.

- 1084 Chao, A.T., Dierick, H.A., Addy, T.M., Bejsovec, A., 2003. Mutations in eukaryotic release
- factors 1 and 3 act as general nonsense suppressors in *Drosophila*. Genetics 165, 601-612.
- 1086 Chen, J., Godt, D., Gunsalus, K., Kiss, I., Goldberg, M., Laski, F.A., 2001. Cofilin/ADF is
- required for cell motility during *Drosophila* ovary development and oogenesis. Nature Cell Biol.
- 1088 3, 204-209.
- 1089 Chen, J.Y., Shen, C., Sivachenko, A.Y., 2006. Mining Alzheimer disease relevant proteins from
- integrated protein interactome data. Pacific Symposium on Biocomputing 11, 367-378.
- 1091 Christiansen, A.E., Ding, T., Bergmann, A., 2012. Ligand-independent activation of the
- Hedgehog pathway displays non-cell autonomous proliferation during eye development in
- 1093 Drosophila. Mech. Dev. 129, 98-108.
- 1094 Cohet, Y., David, J.R., 1978. Control of the Adult reproductive Potential by Preimaginal thermal
- 1095 Conditions. Oecologia 36, 295-306.
- 1096 Couderc, J.L., Godt, D., Zollman, S., Chen, J., Li, M., Tiong, S., Cramton, S.E., Sahut-Barnola,
- 1097 I., Laski, F.A., 2002. The bric a brac locus consists of two paralogous genes encoding BTB/POZ
- domain proteins and acts as a homeotic and morphogenetic regulator of imaginal development
- 1099 in *Drosophila*. Development 129, 2419-2433.
- David, J.R., Bocquet, C., 1975. Similarities and differences in latitudinal adaptation of two
- 1101 *Drosophila* sibling species. Nature 257, 588-590.
- Dobens, L., Jaeger, A., Peterson, J.S., Raftery, L.A., 2005. Bunched sets a boundary for Notch
- signaling to pattern anterior eggshell structures during *Drosophila* oogenesis. Dev. Biol. 287,
- 1104 425-437.
- 1105 Edwards, M.C., Wong, C., Elledge, S.J., 1998. Human cyclin K, a novel RNA polymerase II-
- 1106 associated cyclin possessing both carboxy-terminal domain kinase and Cdk-activating kinase
- 1107 activity. Mol. Cell. Biol. 18, 4291-4300.
- 1108 Foster, L.J., de Hoog, C.L., Zhang, Y., Zhang, Y., Xie, X., Mootha, V.K., Mann, M., 2006. A
- mammalian organelle map by protein correlation profiling. Cell 125, 187-199.
- 1110 Gancz, D., Gilboa, L., 2013. Insulin and Target of rapamycin signaling orchestrate the
- development of ovarian niche-stem cell units in *Drosophila*. Development 140, 4145-4154.
- 1112 Gancz, D., Lengil, T., Gilboa, L., 2011. Coordinated regulation of niche and stem cell precursors
- by hormonal signaling. PLoS Biol. 9, e1001202.
- 1114 Gilboa, L., 2015. Organizing stem cell units in the *Drosophila* ovary. Curr. Op. Genet. Dev. 32C,
- 1115 31-36.
- 1116 Giot, L., Bader, J.S., Brouwer, C., Chaudhuri, A., Kuang, B., Li, Y., Hao, Y.L., Ooi, C.E.,
- 1117 Godwin, B., Vitols, E., Vijayadamodar, G., Pochart, P., Machineni, H., Welsh, M., Kong, Y.,
- 1118 Zerhusen, B., Malcolm, R., Varrone, Z., Collis, A., Minto, M., Burgess, S., McDaniel, L.,
- 1119 Stimpson, E., Spriggs, F., Williams, J., Neurath, K., Ioime, N., Agee, M., Voss, E., Furtak, K.,
- 1120 Renzulli, R., Aanensen, N., Carrolla, S., Bickelhaupt, E., Lazovatsky, Y., DaSilva, A., Zhong, J.,
- 1121 Stanyon, C.A., Finley, R.L., Jr., White, K.P., Braverman, M., Jarvie, T., Gold, S., Leach, M.,

- 1122 Knight, J., Shimkets, R.A., McKenna, M.P., Chant, J., Rothberg, J.M., 2003. A protein
- interaction map of *Drosophila melanogaster*. Science 302, 1727-1736.
- 1124 Godt, D., Couderc, J.L., Cramton, S.E., Laski, F.A., 1993. Pattern formation in the limbs of
- 1125 Drosophila: bric à brac is expressed in both a gradient and a wave-like pattern and is required
- for specification and proper segmentation of the tarsus. Development 119, 799-812.
- 1127 Godt, D., Laski, F.A., 1995. Mechanisms of cell rearrangement and cell recruitment in
- 1128 Drosophila ovary morphogenesis and the requirement of bric à brac. Development 121, 173-
- 1129 187.
- 1130 Gonzalez, G., Uribe, J.C., Tari, L., Brophy, C., Baral, C., 2007. Mining gene-disease
- relationships from biomedical literature: weighting protein-protein interactions and connectivity
- measures. Pacific Symposium on Biocomputing 12, 28-39.
- 1133 Gonzalez, M.W., Kann, M.G., 2012. Chapter 4: Protein interactions and disease. PLoS Comput
- 1134 Biol 8, e1002819.
- 1135 Gramates, L.S., Marygold, S.J., Santos, G.D., Urbano, J.M., Antonazzo, G., Matthews, B.B.,
- 1136 Rey, A.J., Tabone, C.J., Crosby, M.A., Emmert, D.B., Falls, K., Goodman, J.L., Hu, Y., Ponting,
- L., Schroeder, A.J., Strelets, V.B., Thurmond, J., Zhou, P., the FlyBase, C., 2016. FlyBase at
- 1138 25: looking to the future. Nucleic Acids Res. 45, D663-D671.
- 1139 Green II, D.A., Extavour, C.G., 2012. Convergent Evolution of a Reproductive Trait Through
- 1140 Distinct Developmental Mechanisms in *Drosophila*. Dev. Biol. 372, 120-130.
- 1141 Green II, D.A., Sarikaya, D.P., Extavour, C.G., 2011. Counting in oogenesis. Cell Tissue Res.
- 1142 344, 207-212.
- Hahn, M.W., Kern, A.D., 2005. Comparative genomics of centrality and essentiality in three
- 1144 eukaryotic protein-interaction networks. Mol. Biol. Evol. 22, 803-806.
- Harris, T.J., Peifer, M., 2004. Adherens junction-dependent and -independent steps in the
- 1146 establishment of epithelial cell polarity in *Drosophila*. J. Cell Biol. 167, 135-147.
- 1147 Hartwell, L.H., Hopfield, J.J., Leibler, S., Murray, A.W., 1999. From molecular to modular cell
- 1148 biology. Nature 402, C47-C52.
- Hilman, D., Gat, U., 2011. The evolutionary history of YAP and the hippo/YAP pathway. Mol.
- 1150 Biol. Evol. 28, 2403-2417.
- Hodin, J., Riddiford, L.M., 1998. The ecdysone receptor and ultraspiracle regulate the timing
- and progression of ovarian morphogenesis during *Drosophila* metamorphosis. Dev. Genes Evol.
- 1153 208, 304-317.
- Hodin, J., Riddiford, L.M., 2000a. Different mechanisms underlie phenotypic plasticity and
- interspecific variation for a reproductive character in Drosophilids (Insecta: Diptera). Evolution 5,
- 1156 1638-1653.

- Hodin, J., Riddiford, L.M., 2000b. Parallel alterations in the timing of ovarian ecdysone receptor
- and ultraspiracle expression characterize the independent evolution of larval reproduction in two
- 1159 species of gall midges (Diptera: Cecidomyiidae). Dev. Genes Evol. 210, 358-372.
- 1160 Hsu, H.-J., Drummond-Barbosa, D., 2009. Insulin levels control female germline stem cell
- maintenance via the niche in *Drosophila*. Proc. Natl. Acad. Sci. USA 106, 1117-1121.
- Huang, S.S., Fraenkel, E., 2009. Integrating proteomic, transcriptional, and interactome data
- reveals hidden components of signaling and regulatory networks. Science Signaling 2, ra40.
- 1164 Ideker, T., Sharan, R., 2008. Protein networks in disease. Genome Res. 18, 644-652.
- 1165 Ito, T., Bulger, M., Kobayashi, R., Kadonaga, J.T., 1996. *Drosophila* NAP-1 is a core histone
- chaperone that functions in ATP-facilitated assembly of regularly spaced nucleosomal arrays.
- 1167 Mol. Cell. Biol. 16, 3112-3124.
- Jalili, M., Salehzadeh-Yazdi, A., Gupta, S., Wolkenhauer, O., Yaghmaie, M., Resendis-Antonio,
- 1169 O., Alimoghaddam, K., 2016. Evolution of Centrality Measurements for the Detection of
- 1170 Essential Proteins in Biological Networks. Frontiers in Physiology 7, 375.
- Jansen, R., Greenbaum, D., Gerstein, M., 2002. Relating whole-genome expression data with
- protein-protein interactions. Genome Res. 12, 37-46.
- 1173 Jeong, H., Mason, S.P., Barabási, A.L., Oltvai, Z.N., 2001. Lethality and centrality in protein
- 1174 networks. Nature 411, 41-42.
- 1175 Jiang, K., Liu, Y., Zhang, J., Jia, J., 2018. An intracellular activation of Smoothened that is
- independent of Hedgehog stimulation in *Drosophila*. J. Cell Sci. 131, jcs211367.
- 1177 Kambysellis, M.P., Heed, W.B., 1971. Studies of Oogenesis in Natural Populations of
- 1178 Drosophilidae. I. Relation of ovarian development and ecological habitats of the Hawaiian
- 1179 species. Am. Nat. 941, 31-49.
- 1180 Kanehisa, M., Goto, S., Furumichi, M., Tanabe, M., Hirakawa, M., 2010. KEGG for
- 1181 representation and analysis of molecular networks involving diseases and drugs. Nucleic Acids
- 1182 Res. 38, D355-360.
- Kasza, K.E., Farrell, D.L., Zallen, J.A., 2014. Spatiotemporal control of epithelial remodeling by
- regulated myosin phosphorylation. Proc. Natl. Acad. Sci. USA 111, 11732-11737.
- Keller, R., 2006. Mechanisms of elongation in embryogenesis. Development 133, 2291-2302.
- 1186 King, R.C., 1970. Ovarian Development in *Drosophila melanogaster*. Academic Press, New
- 1187 York.
- King, R.C., Aggarwal, S.K., Aggarwal, U., 1968. The Development of the Female *Drosophila*
- 1189 Reproductive System. J. Morphol. 124, 143-166.
- 1190 Klepsatel, P., Galikova, M., De Maio, N., Huber, C.D., Schlotterer, C., Flatt, T., 2013a. Variation
- in thermal performance and reaction norms among populations of *Drosophila melanogaster*.
- 1192 Evolution 67, 3573-3587.

- Klepsatel, P., Galikova, M., De Maio, N., Ricci, S., Schlotterer, C., Flatt, T., 2013b. Reproductive
- and post-reproductive life history of wild-caught *Drosophila melanogaster* under laboratory
- 1195 conditions. J. Evol. Biol. 26, 1508-1520.
- 1196 Koschutzki, D., Schreiber, F., 2008. Centrality analysis methods for biological networks and
- their application to gene regulatory networks. Gene Regulation and Systems Biology 2, 193-
- 1198 201.
- 1199 LaFever, L., Drummond-Barbosa, D., 2005. Direct control of germline stem cell division and cyst
- growth by neural insulin in *Drosophila*. Science 309, 1071-1073.
- 1201 LaFever, L., Feoktistov, A., Hsu, H.J., Drummond-Barbosa, D., 2010. Specific roles of Target of
- rapamycin in the control of stem cells and their progeny in the *Drosophila* ovary. Development
- 1203 137, 2117-2126.
- Lai, C.M., Lin, K.Y., Kao, S.H., Chen, Y.N., Huang, F., Hsu, H.J., 2017. Hedgehog signaling
- establishes precursors for germline stem cell niches by regulating cell adhesion. J. Cell Biol.
- 1206 216, 1439-1453.
- Lee, R.T., Zhao, Z., Ingham, P.W., 2016. Hedgehog signalling. Development 143, 367-372.
- 1208 Lewellyn, L., Cetera, M., Horne-Badovinac, S., 2013. Misshapen decreases integrin levels to
- promote epithelial motility and planar polarity in *Drosophila*. J. Cell Biol. 200, 721-729.
- 1210 Li, X., Wu, M., Kwoh, C.K., Ng, S.K., 2010. Computational approaches for detecting protein
- 1211 complexes from protein interaction networks: a survey. BMC Genomics 11 Suppl 1, S3.
- 1212 Llamazares, S., Moreira, A., Tavares, A., Girdham, C., Spruce, B.A., Gonzalez, C., Karess,
- 1213 R.E., Glover, D.M., Sunkel, C.E., 1991. polo encodes a protein kinase homolog required for
- mitosis in *Drosophila*. Genes Dev. 5, 2153-2165.
- Lobell, A.S., Kaspari, R.R., Serrano Negron, Y.L., Harbison, S.T., 2017. The Genetic
- 1216 Architecture of Ovariole Number in *Drosophila melanogaster*. Genes with Major, Quantitative,
- 1217 and Pleiotropic Effects. G3 7, 2391-2403.
- 1218 Ma'ayan, A., 2011. Introduction to network analysis in systems biology. Sci Signal 4, tr5.
- 1219 Markow, T.A., Beall, S., Matzkin, L.M., 2009. Egg size, embryonic development time and
- ovoviviparity in *Drosophila* species. J. Evol. Biol. 22, 430-434.
- Matthews, L.R., Vaglio, P., Reboul, J., Ge, H., Davis, B.P., Garrels, J., Vincent, S., Vidal, M.,
- 1222 2001. Identification of potential interaction networks using sequence-based searches for
- 1223 conserved protein-protein interactions or "interologs". Genome Res. 11, 2120-2126.
- 1224 Mbodj, A., Junion, G., Brun, C., Furlong, E.E., Thieffry, D., 2013. Logical modelling of
- 1225 Drosophila signalling pathways. Molecular BioSystems 9, 2248-2258.
- 1226 Mizuno, T., Amano, M., Kaibuchi, K., Nishida, Y., 1999. Identification and characterization of
- 1227 Drosophila homolog of Rho-kinase. Gene 238, 437-444.

- 1228 Orgogozo, V., Broman, K.W., Stern, D.L., 2006. High-resolution quantitative trait locus mapping
- reveals sign epistasis controlling ovariole number between two *Drosophila* species. Genetics
- 1230 173, 197-205.
- 1231 R'Kha, S., Capy, P., David, J.R., 1991. Host-plant specialization in the *Drosophila melanogaster*
- 1232 species complex: A physiological, behavioral, and genetic analysis. Proc. Natl. Acad. Sci. USA
- 1233 88, 1835-1839.
- 1234 R'kha, S., Moreteau, B., Coyne, J.A., David, J.R., 1997. Evolution of a lesser fitness trait: egg
- production in the specialist *Drosophila sechellia*. Genetical research 69, 17-23.
- 1236 Ravasz, E., Somera, A.L., Mongru, D.A., Oltvai, Z.N., Barabási, A.L., 2002. Hierarchical
- organization of modularity in metabolic networks. Science 297, 1551-1555.
- 1238 Ridley, A.J., 2006. Rho GTPases and actin dynamics in membrane protrusions and vesicle
- 1239 trafficking. Trends Cell Biol. 16, 522-529.
- 1240 Sahut-Barnola, I., Dastugue, B., Couderc, J.-L., 1996. Terminal filament cell organization in the
- larval ovary of *Drosophila melanogaster*: ultrastructural observations and pattern of divisions.
- 1242 Roux's Archives of Developmental Biology 205, 356-363.
- Sahut-Barnola, I., Godt, D., Laski, F.A., Couderc, J.-L., 1995. *Drosophila* Ovary Morphogenesis:
- 1244 Analysis of Terminal Filament Formation and Identification of a Gene Required for This Process.
- 1245 Dev. Biol. 170, 127-135.
- Sarikaya, D.P., Belay, A.A., Ahuja, A., Green II, D.A., Dorta, A., Extavour, C.G., 2012. The roles
- of cell size and cell number in determining ovariole number in *Drosophila*. Dev. Biol. 363, 279-
- 1248 289
- 1249 Sarikaya, D.P., Church, S.H., Lagomarsino, L.P., Magnacca, K.M., Montgomery, S.L., Price,
- 1250 D.P., Kaneshiro, K.Y., Extavour, C.G., 2019. Reproductive capacity evolves in response to
- ecology through common developmental mechanisms in Hawai'ian *Drosophila*. Curr. Biol. 29,
- 1252 1877-1884.
- 1253 Sarikaya, D.P., Extavour, C.G., 2015. The Hippo pathway regulates homeostatic growth of stem
- 1254 cell niche precursors in the *Drosophila* ovary. PLoS Genetics 11, e1004962.
- 1255 Sebe-Pedros, A., Zheng, Y., Ruiz-Trillo, I., Pan, D., 2012. Premetazoan origin of the hippo
- 1256 signaling pathway. Cell Reports 1, 13-20.
- 1257 Spellman, P.T., Sherlock, G., Zhang, M.Q., Iyer, V.R., Anders, K., Eisen, M.B., Brown, P.O.,
- Botstein, D., Futcher, B., 1998. Comprehensive identification of cell cycle-regulated genes of the
- 1259 yeast Saccharomyces cerevisiae by microarray hybridization. Mol. Biol. Cell 9, 3273-3297.
- 1260 Srinivasan, B.S., Shah, N.H., Flannick, J.A., Abeliuk, E., Novak, A.F., Batzoglou, S., 2007.
- 1261 Current progress in network research: toward reference networks for key model organisms.
- 1262 Brief. Bioinform. 8, 318-332.
- von Mering, C., Krause, R., Snel, B., Cornell, M., Oliver, S.G., Fields, S., Bork, P., 2002.
- 1264 Comparative assessment of large-scale data sets of protein-protein interactions. Nature 417,
- 1265 399-403.

- 1266 Wachi, S., Yoneda, K., Wu, R., 2005. Interactome-transcriptome analysis reveals the high
- centrality of genes differentially expressed in lung cancer tissues. Bioinformatics 21, 4205-4208.
- Wang, R.S., Hall, K.T., Giulianini, F., Passow, D., Kaptchuk, T.J., Loscalzo, J., 2017. Network
- analysis of the genomic basis of the placebo effect. JCI Insight 2, 93911.
- 1270 Wang, R.S., Loscalzo, J., 2018. Network-Based Disease Module Discovery by a Novel Seed
- 1271 Connector Algorithm with Pathobiological Implications. J. Mol. Biol. 430, 2939-2950.
- 1272 Watts, D.J., Strogatz, S.H., 1998. Collective dynamics of 'small-world' networks. Nature 393,
- 1273 440-442.
- Wayne, M.L., Hackett, J.B., Dilda, C.L., Nuzhdin, S.V., Pasyukova, E.G., Mackay, T.F., 2001.
- 1275 Quantitative trait locus mapping of fitness-related traits in *Drosophila melanogaster*. Genetical
- 1276 research 77, 107-116.
- 1277 Wayne, M.L., Hackett, J.B., Mackay, T.F.C., 1997. Quantitative Genetics of Ovariole Number in
- 1278 Drosophila melanogaster. I. Segregating Variation and Fitness. Evolution 4, 1156-1163.
- 1279 Wayne, M.L., McIntyre, L.M., 2002. Combining mapping and arraying: An approach to candidate
- gene identification. Proc. Natl. Acad. Sci. USA 99, 14903-14906.
- 1281 Yang, X., Xu, T., 2011. Molecular mechanism of size control in development and human
- 1282 diseases. Cell Research 21, 715-729.
- 1283 Yook, S.H., Oltvai, Z.N., Barabási, A.L., 2004. Functional and topological characterization of
- 1284 protein interaction networks. Proteomics 4, 928-942.
- 1285 Yu, H., Paccanaro, A., Trifonov, V., Gerstein, M., 2006. Predicting interactions in protein
- networks by completing defective cliques. Bioinformatics 22, 823-829.
- 1287 Zhang, Y., Lin, H., Yang, Z., Wang, J., Liu, Y., 2017. An uncertain model-based approach for
- 1288 identifying dynamic protein complexes in uncertain protein-protein interaction networks. BMC
- 1289 Genomics 18, 743.

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